Short Communication:

Control of anthracnose in "Kluai Hom Tong" banana (Musa (AAA group)) using mangosteen pericarp crude extract

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ABSTRACT

Anthracnose is a postharvest disease caused by the fungus Colletotrichum musae that results in major economic losses during transportation and storage of bananas. Mangosteen fruit has been widely used as traditional medicine and also as an antimicrobial agent against plant pathogenic fungi. The effectiveness of mangosteen pericarp crude extract on controlling anthracnose in "Kluai Hom Tong" banana (Musa (AAA group)) was evaluated. The fruits were dipped in various concentrations (0, 1,000, 1,200 and 1,400 mg L⁻¹) of mangosteen pericarp methanol crude extract solution for 2 min and then sprayed with 1 mL of 10⁶ spores mL⁻¹ of Colletotrichum musae. Incubated fruits at 29 ± 2°C for 2 days and stored at 13 ± 2°C for 14 days. Banana fruits were further directly sprayed with 1 mL of 500 mg L⁻¹ of ethephon solution and incubated for one day at room temperature. Anthracnose disease percentage and quality of banana fruits as total soluble solids (TSS), titratable acid (%TA), fresh weight and color were recorded. Color measurements were recorded 2 days interval. The results showed that mangosteen pericarp crude extract treatment demonstrated a lower disease percentage than non-treated fruits and the lowest disease percentage was found in the 1,200 mg L⁻¹ treatment at 45% with 8 days of shelf life.

Keywords: Mangosteen, Musa (AAA group), postharvest disease, Colletotrichum musae

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INTRODUCTION

Bananas are an important agricultural commodity in Thailand and are grown throughout the country (National Committee on Agricultural Commodity and Food Standards, 2008). The main variety of bananas is the "Hom" banana, especially the "Hom Thong" variety (Anuchai et al., 2018). In 2012, Thailand produced "Kluai Hom Tong" banana (Musa (AAA group)) at 231,031 Mt for domestic use and 2,169 Mt for export (OAE, 2012). Worldwide, postharvest losses of fruits and vegetables could average about 45% (Kitinoja and Kader, 2015). One limiting factor that influences the economic value of banana is its relatively short shelf-life caused by postharvest pathogen attack (Kuyu and Tola, 2018). The anthracnose is a postharvest disease of banana that can result in 30-40% losses of marketable fruit during transportation and storage (Ranasinghe et al., 2003). Anthracnose is caused by the fungus Colletotrichum musae (Jeffries et al., 1990). Antimicrobial activities in plant extracts occur in both in vitro and in vivo fungal development. Cruz et al. (2013) discovered that extracts from Allium sativum,



Copaifera langsdorffii. Cinnamomum zevlanicum and Eugenia caryophyllata gave effective control of postharvest anthracnose of banana fruit while Montri et al. (2010) found that Stemona curtisii Hook, f. crude extract inhibited the fungus C. musae and reduced anthracnose disease in "Kluai Hom Tong" banana.

Mangosteen is a tropical fruit from the family Guttiferae. Thailand has high annual mangosteen production at approximately three million Mt. Mangosteen pericarp is the largest part of the fruit. comprising 66.67% of the total fruit (Siripanich and Luckanatinvong, 1997). The pericarp is usually discarded as waste due to its unpleasant bitter taste (Pedraza-Chaverri et al., 2008). However, mangosteen pericarp has long been used as a traditional indigenous medicine across Southeast Asia, especially for anti-inflammatory (Chomnawang et al., 2005; Akao et al., 2008), anti-oxidant (Jung et al., 2006), anti-proliferative (Pedraza-Chaverri et al., 2008) and immunostimulatory biological activities (Matsumoto et al., 2003). It has also been used as an antimicrobial (Al-Massarani et al., 2013; Palakawong et al., 2013) against plant pathogenic fungi. Sangsuwan et al. (2011) reported that mangosteen pericarp powder inhibited anthracnose fungi in "Kluai Hom Tong" banana. The use of waste for the production of various crucial bioactive components is an important step toward sustainable development (Sagar et al., 2018). Here, the effectiveness of mangosteen pericarp crude extract was evaluated for the control of anthracnose to extend shelf life in free toxin "Kluai Hom Tong".

MATERIALS AND METHODS

Raw Material

Toxic-free "Kluai Hom Tong" banana (Musa (AAA group)) fruits at the 70% mature stage were purchased from Thung Kha Wat farmers, Lamae District, Chumphon Province to avoid bruising and injury from the harvesting process. The fruit bunches were de-handed and selected for fruit weight (110 ± 10 g) with uniformity of shape and size. The crown portions were wrapped up to the mid-portion

of the hand with paper to prevent abrasion and latex staining of other fruits. Fruits were cleaned following the process to export free toxin "Kluai Hom Tong" bananas to Japan by removal of hull and dirt of sap, removal of insect, sap washing after cutting, cleaning the fruits with tap-water and air-drying at room temperature.

Isolation of the Pathogen and Inoculum Preparation

Banana fruits showing anthracnose disease symptoms were collected from the "Kluai Hom Tong" banana orchard in Chumphon and used to isolate Colletotrichum musae. The pure culture was maintained on potato dextrose agar (PDA) medium. The inoculum was prepared by flooding the surface of 8 days old culture with sterile distilled water, scraping the surface gently with a glass rod. Spores were counted using a hemacytometer (Hausser Scientific, Horsham, PA, USA) and the spore concentration was adjusted to 106 spores mL^{-1} .

Mangosteen Pericarp Methanolic Crude Extract

Mangosteen fruits at maturity stage 4-5 were bought from a farmer in Lang Suan District, Chumphon Province, Thailand. Fruits were stored at the ambient temperature till the color turns to stage 6 or when the skin color turns to blackish purple. The crude extract was prepared by cutting the fruit pericarp into small pieces. Pieces of fruit pericarp were dried in a hot air oven at 60°C for 24 h. then ground by using a blender. The dried powder was macerated in 95% methanol for 72 h (3 times) and the extracted was concentrated by using a rotary evaporator. A stock solution of crude extract was prepared at the concentration of 10,000 mg L⁻¹ and kept in the refrigerator.

Treatments and Applications of Mangosteen Pericarp Extract

The experiment was conducted in a completely randomized design (CRD) with 4 treatments and 10 replications. A cluster of three fingers was considered as a replicate. After

cleaning and drving, the fruits were dipped into various concentrations of mangosteen pericarp crude methanolic extract solutions at the concentrations of 0, 1,000, 1,200 and 1,400 mg L⁻¹ for 2 min then sprayed with 1 mL of 10⁶ spores mL⁻¹ of Colletotrichum musae spore suspension. incubated for 2 days at room temperature (29 ± 2°C) for fungal growth, and following the transportation process to Japan, the fruit clusters were stored in the control room at 13 ± 2°C for 14 days. Banana fruits were further directly sprayed with 1 mL of 500 mg L⁻¹ of ethephon solution and incubated for 1 day at room temperature. Anthracnose disease in each fruit was measured in 2 days interval for incidence and lesion. Quality of banana fruits as total soluble solids (TSS), titratable acid (%TA), fresh weight and color were recorded. The percentage of weight loss and disease incidence were calculated.

Weight Loss (%)

Banana fruits were weighed before and after storage at an interval of 2 days. The percentage of weight loss was calculated using the expression given below.

Weight loss (%) = Initial weight (g) - Weight (g) at sampling date / Initial weight (g) × 100

Color Measurement

Peel and pulp color of banana fruits were measured using a HunterLab MiniScan Photometer (MiniScan@ XE Plus; Hunter Associates Laboratory Inc.; Reston, VA, USA). The CIELAB color scale which is lightness (L*), greenness (-a*), redness (+a*) and yellowness (b*) values were recorded (Boun and Huxsoll, 1991). The color determination was performed at the top, middle and bottom regions from three different faces of an individual finger.

Firmness Measurement

Three fingers from each hand were used for firmness measurement. The peeled banana was placed in a Texture Analyzer (TA Plus; Lloyd Limited; Fareham, UK) with a 1 mm cylindrical probe at the middle part of the fruit. The firmness value was recorded in Newtons and reported as mean of three readings.

Total Soluble Solids (%) and Total Acidity Measurements

Ten grams of banana pulp tissue were homogenized with 30 mL of distilled water and filtered. Total soluble solids of banana fruit pulp were measured with a refractometer (Model no. ATC-1090, Atago Co., Ltd., Tokyo, Japan) with readings expressed as a percentage. Readings were recorded by pouring two drops of filtrate on the glass prism of the refractometer. Then, a 5 mL aliquot was titrated with 0.1 N NaOH using 1% (w/v) phenolphthalein as an indicator, the results were expressed as a percentage of malic acid in flesh pulp (AOAC, 1990).

Statistical Analysis

All data analyses were carried out in ten replications and results were submitted for analysis of variance (ANOVA) with a significance level of 95% (P < 0.05).

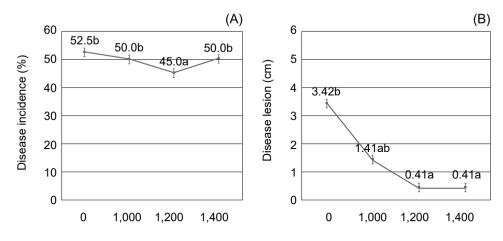
RESULTS AND DISCUSSION

Results showed that mangosteen pericarp crude methanolic extracts had no effect on TSS. TA. pulp firmness and weight loss percentage, however represented potential inhibitory activity against Colletotrichum musae (Table 1). Lower disease incidence percentage and fewer disease lesions were achieved in treated fruits compared with non-treated fruits as control. The lowest disease incidence of 45% was found at the concentration of 1,200 mg L⁻¹ treatment. The disease incidence percentage was high when using a higher concentration of crude extract at the concentration of 1,400 mg L⁻¹ (Figure 1). The quality of ripening banana fruits was controlled by ethylene treatment in commercial practices. During banana ripening, peel color changes from green to yellow, the flavor develops and the pulp softens. However, the results showed that



non-treated fruits with shelf life of 4 days improved to 6 days for mangosteen pericarp methanolic crude extract treated fruits at the concentrations of 1,000–1,200 ppm (Figure 2). The quality of

banana fruit also improved in peel firmness (Table 1) and peel color (Table 2) at the concentration of 1,000 mg L^{-1} treatment, with 8 days of shelf life.



Mangosteen pericarp crude methanolic extract (mgL-1)

Figure 1 Disease incidence percentage (A) and disease lesions (B) of treated banana fruits with various concentrations of mangosteen pericarp crude methanolic extract before spraying with spore suspension of *Colletotrichum musae* at 8 days of shelf life

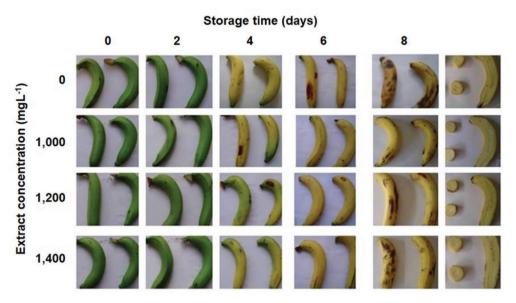


Figure 2 External and internal appearances of banana fruits after soaking with different concentrations of mangosteen pericarp crude methanolic extract before spraying with spore suspension of *Colletotrichum musae*

Extracts from mangosteen peels contain various secondary metabolites such as polyphenols (Moosophin et al., 2010) and prenylated and oxygenated xanthones (Ji et al., 2007). Disease incidence and quality were improved by several active compounds contained in mangosteen peel extract with 95% ethanol as xanthones, flavonoids. tannins, terpenoids and saponins (Saepudin et al., 2019). Several studies showed that xanthones from mangosteen have antibacterial, antifungal and antiviral biological activities (Vishnu et al., 2010: Al-Massarani et al., 2013; Saepudin et al., 2019). Tannins and flavonoids are a class of complex molecules of polyphenols (Saepudin et al., 2019). They can slow down and inhibit the growth of many fungi, yeasts, bacteria and viruses (Kaka et al., 2019), while tannins inhibit the protein transport enzymes through cell membranes. Sangsuwan et al. (2011) reported that fruits coated with mangosteen pericarp powder inhibited anthracnose in "Kluai Hom Tong" banana (Musa (AAA group)). Moreover, antifungal activity of the extract from Garcinia mangostana against the three phytopathogenic fungi Fusarium oxysporum. Alternaria tenuis and Drechslera orvzae was reported by Gopalakrishnan et al. (1997). Our results indicated that mangosteen pericarp crude extract showing the effectiveness of disease control and fruit quality of "Kluai Hom Tong" banana (Musa (AAA group)). However, disease incidence percentage was higher when increased the concentrations of the crude extract due to the negative effect resulting from enhanced total polyphenols (Afifah and Niwat, 2015).

Table 1 Total soluble solids (TSS), titratable acid (%TA), peel and pulp firmness and weight loss percentage of treated banana fruits with various concentrations of mangosteen pericarp crude methanolic extract before spraying with spore suspension of Colletotrichum musae at 8 days of shelf life

Crude extract (mg L ⁻¹)	TSS (%)	TA (%)	Peel firmness (N)	Pulp firmness (N)	Weight Loss (%)	
0	5.75	0.36	14.94 ^b	5.23	5.50	
1,000	6.30	0.32	19.74ª	5.71	6.36	
1,200	6.75	0.31	16.07 ^b	5.97	6.15	
1,400	6.60	0.34	15.10⁵	6.23	5.93	
F-test	ns	ns	*	ns	ns	
CV (%)	14.02	12.76	18.94	16.85	67.90	

Note: CV = coefficient of variation, mean values in columns followed by the same small letter are not statistically different using Duncan's test at 5% probability, ns = non-significant, * P < 0.05



Table 2 Peel and pulp color (L*, a* and b*) of treated banana fruits with various concentrations of mangosteen pericarp methanolic crude extract before spraying with spore suspension of *Colletotrichum musae* at 8 days of shelf life

Crude extract (mg L⁻¹)	Peel color			Pulp color		
	L*	a*	b*	L*	a*	b*
0	63.68ª	43.97	10.73	75.73ª	26.65b	2.21b
1,000	52.00b	35.24	11.69	75.38ª	26.81 ^b	2.47ab
1,200	57.90 ^{ab}	38.69	10.97	73.05 ^b	30.60a	2.84ª
1,400	53.03b	35.84	11.27	76.07ª	23.64°	1.96 ^b
F-test	*	ns	ns	*	*	*
CV (%)	14.75	20.91	11.46	1.35	8.86	20.01

Note: CV = coefficient of variation, mean values in columns followed by the same small letter are not statistically different using Duncan's test at 5% probability, ns = non-significant, * P < 0.05

CONCLUSIONS

ACKNOWLEDGEMENTS

Our results suggested that crude methanolic extracts of mangosteen pericarp were effective for disease control of banana fruit. Treatments preserved acceptable and better peel color with a lower incidence of anthracnose disease

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REFERENCES

- Afifah, R.A. and C. Niwat. 2015. The use of mangosteen pericarp (*Garcinia mangostana* L.) extract to fortify the green tea drink enchanced antioxidant activity. KKU Res. J. 20(3): 305–313.
- Akao, Y., Y. Nakagawa, M. linuma and Y. Nozawa. 2008. Anti-cancer effects of xanthones from pericarps of mangosteen. Int. J. Mol. 9: 355–370.
- Al-Massarani, S.M., A.A. El Gamal, N.M. Al-Musayeib, R.A. Mothana, O.A. Basudan, A.J. Al-Rehaily, M. Farag, M.H. Assaf, K.H. El Tahir and L. Maes. 2013. Phytochemical, antimicrobial and antiprotozoal evaluation of *Garcinia mangostana* pericarp and α-mangostin, its major xanthone derivative. Molecules 18: 10599–10608.
- Anuchai, J., M. Chumthongwattana, R. Tepsorn and S. Supapavnich. 2018. Efficiency of salicylic acid immersion using fine-bubble technique on quality of *Musa* AAA fruit during ripening. IJAT. 14(7): 1003–1016.
- AOAC (Association of Official Analytical Chemists). 1990. Official Methods of Analysis. 15th edition. Washington, D.C., USA.
- Boun, H.R. and C.C. Huxsoll. 1991. Control of minimally processed carrot (*Daucus carota*) surface discoloration caused by abrasion peeling. J. Food Sci. 56: 416–418.

- Chomnawang, M.T., S. Surassmo, V.S. Ukoolkarn and W. Gritsanapan, 2005, Antimicrobial effects of Thai medicinal plants against agne-inducing bacteria. J. Ethnopharmacol. 3(101): 330–333.
- Cruz, M.E.S., K.R.F. Schwan-estrada, E. Clemente, A.T. Itako, J.R. Stangarlin and M.J.S. Cruz, 2013. Plant extracts for controlling the postharvest anthracnose of banana fruit. Rev. Bras. Plantas Med. 15(4)(Suppl. I): 727-733.
- Gopalakrishnan, G., B. Banumathi and G. Suresh, 1997, Evaluation of the antifungal of natural xanthones from Garcinia mangostana and their synthetic derivatives, J. Nat. Prod. 60: 519-524.
- Jeffries, P., J.C. Dodd, M.J. Jeger and R.A. Plumblev, 1990, The biology and control of Colletotrichum species on tropical fruit crops. J. Plant Pathol. 39: 343-366.
- Ji, X., B. Avula and I.A. Khan, 2007. Quantitative and qualitative determination of six xanthones in Garcinia mangostana L. by LC-PDA and LC-ESI-MS. J. Pharm. Biomed. Anal. 43(4): 1270–1276.
- Jung, H.A., B.N. Su, W.J. Keller and A.D. Kinghorn. 2006. Antioxidant Xanthones from the pericarp of Garcinia Mangostana (Mangosteen). J. Agric. Food Chem. 54: 2077–2082.
- Kaka, A.K., K.A. Ibupoto, S.H. Chattha, S.A. Soomro, H. Mangio, S.A. Juneio, A.H. Soomro, S.G. Khaskheli and S.K. Kaka, 2019, Effect of hot water treatments and storage period on the quality attributes of banana (Musa sp.) fruit. Pure Appl. Biol. 8(1): 363–371.
- Kitinoja, L. and A.A. Kader. 2015. Measuring Postharvest Losses of Fresh Fruits and Vegetables in Developing Countries, PEF White Paper 15–02. The Postharvest Education Foundation. Oregon, USA.
- Kuyu, G.C. and Y.B. Tola. 2018. Assessment of banana fruit handling practices and associated fungal pathogens in Jimma town market, southwest Ethiopia. Food Sci. Nutr. 6(3): 609-616.
- Matsumoto, K., Y. Akao, E. Kobayashi, K. Ohguchi, T. Ito, T. Tanaka, M. Iinuma and Y. Nozawa. 2003. Induction of apoptosis by xanthones from mangosteen in human leukemia cell lines. J. Nat. Prod. 66(8): 1124-1127.
- Montri, N., K. Bunya-atichart and P. Leuiantuak. 2010. Effect of crude extract from Stemona curtisii Hook.f. on anthracnose controlling in 'Kluai Hom Tong' Musa sp. (AAA group). Agri. Sci. J. 41(1)(Suppl.): 333-336.
- Moosophin, K., T. Wetthaisong, L. Seeratchakot and W. Kokluecha. 2010. Tannin extraction from mangosteen peel for protein precipitation in wine. KKU Res. J. 15(5): 377-385.
- National Committee on Agricultural Commodity and Food Standards. 2008. Available Source: http:// www.acfs.go.th/standard/download/eng/bananas.pdf.
- OAE (Office of Agricultural Economics). 2012. Available Source: http://www.oae.go.th/.
- Palakawong, C., P. Sophanodora, P. Toivonen and P. Delaguis. 2013. Optimized extraction and characterization of antimicrobial phenolic compounds from mangosteen (Garcinia mangostana L.) cultivation and processing waste. J. Sci. Food Agric. 93: 3792–3800.
- Pedraza-Chaverri, J., N.C. Rodríguez, M.J. Orozco-Ibarra and M. Pérez-Rojas. 2008. Medicinal properties of mangosteen (Garcinia mangostana). Food Chem. Toxicol. 46: 3227-3239.
- Ranasinghe, L.S., B. Jayawardena and K. Abeywickrama. 2003. Use of waste generated from cinnamon bark oil extraction as a postharvest treatment of Embul banana. J. Food Agric. Environ. 1(1): 340-344.



- Saepudin, A., D. Natawijaya, E. Hartini and R. Iskandar. 2019. Evaluation of antibacterial activity of mangosteen (*Garcinia mangostana* L.) pericarp extract against rice leaf blight bacteria (*Xanthomonas oryzae* pv. oryzae) at various temperatures and durations of fruit storage. IOP Conference Series: Earth Environmental Science. 250: 012026.
- Sagar, N.A., S. Pareek, S. Sharma, E.M. Yahia and M.G. Lobo. 2018. Fruit and vegetable waste: bioactive compounds, their extraction, and possible utilization. Compr. Rev. Food Sci. F. 17(3): 512–531.
- Sangsuwan, J., S. Khamthai and P. Thongchai. 2011. Utilization of mangosteen pericarp powder for anti–anthracnose disease packaging material production in "Hom Thong" banana (*Musa* AAA group). Agri. Sci. J. 42(1)(Suppl.): 579–582.
- Siripanich, J. and V. Luckanatinvong. 1997. Chemical composition and the development of flesh translucent disorder in mangosteen. *In*: Proceedings of The Australian Postharvest Holticulture Conference. Australia, Sydney.
- Vishnu, P.V., M. Jainu, S.K. Mohan, P. Saraswathi and V.S.C. Gopan. 2010. Antimicrobial activity of pericarp extract of *Garcinia mangostana* Linn. Int. J. Pharm. Sci. Rev. Res. 1(8): 278–281.