

EFFECTS OF THE ESSENTIAL OILS OF IRANIAN MEDICINAL PLANTS ON THE GROWTH OF SOME PATHOGENIC BACTERIA

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ABSTRACT

The antimicrobial activities of the essential oils of four different medicinal plants (cumin, thyme, mint and fennel) were investigated on *Escherichia coli*, *Klebsiella pneumonia*, *Proteus vulgaris*, and *Pseudomonas aeruginosa*. The antimicrobial activity of the essential oils was evaluated by disc diffusion method. The results showed that the most effective oils were thyme (*Thymus vulgaris* L.) and peppermint (*Mintae piperita*). The use of these essential oils, should be considered in the food preservation.

KEYWORDS: Essential oil, *Escherichia coli*, *Proteus vulgaris*, *Klebsiella pneumoniae*, *Pseudomonas aeruginosa*, Antibacterial activity

1. INTRODUCTION

The interest in the substitution of traditional food preservatives, as antimicrobial and antioxidants, by natural compounds has fostered research on vegetable sources and the screening of plant materials in order to identify new compounds. It has been known since ancient times that medicinal, aromatic and spice plants and their essential oils (active substances) have varying degrees of antimicrobial activities [1-14]. The major antimicrobial substances of these plants and their essential oils are, for example, eugenol in cloves, allicin in garlic, cinnamic aldehyde and eugenol in cinnamon, carvacrol and thymol in oregano and thyme, and vanillin in vanilla beans [14-15]. Furthermore, essential oils of many aromatic plants have shown to possess antimicrobial activities [16-17]. For example, oils from basil, bay, clove, thyme and rosemary showed bactericidal activities against *L. monocytogenesis* and other pathogens [18-19]. Incidences of foodborne illnesses are still a major problem, even in developed countries. In fact, food poisoning is still a threat for both consumers and the food industry despite the use of preserving processes. Meanwhile, consumers are concerned about the safety of foods containing preservatives. Therefore, there has been a growing interest in new and effective techniques to reduce cases of food borne illnesses. There is considerable

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interest in the possible use of these compounds as food additives, to delay the onset of food spoilage or to prevent the growth of food borne pathogens.

The aim of the present study was to evaluate the antibacterial activity of some edible and medicinal plant essential oils against common pathogens.

2. MATERIALS AND METHODS

2.1 Extraction of essential oil

The plants investigated in this study are listed in Table 1. The essential oils were extracted by Clevenger instrument, via water distillation. The plant material was placed in water, heated to a boil and the steam carrying the essential oil is condensed and collected in a receiver flask and then essential oils separated from water by separator container.

2.2 Microorganisms

Escherichia coli, *Klebsiella pneumoniae*, *Proteus vulgaris*, *Pseudomonas aeruginosa* were used as test organisms. The bacteria were incubated on nutrient agar slants for 48 h at 37°C followed by inoculation in Mueller Hinton Agar medium. The bacteria were supplied by the Department of Microbiology, Medical Science university, Mashhad, Iran.

2.3 Antibacterial assay

Antibacterial activity was demonstrated using a modification of the method originally described by Kirby-Bauer (1966) which is widely used for the antibacterial susceptibility testing [20]. A loopful bacteria was taken from the stock culture and dissolved in 1 ml of brain heart infusion broth (BHI) and incubated at 35 °C. Final cell concentration were 10^6 - 10^7 cfu/ml. One ml of this inoculum was added to each plate containing nutrient agar. All the tests were done by placing the paper discs (50 μ l for 5 mm diameter filter paper disc) with various concentrations of essential oils on the Mueller Hinton Agar surface previously inoculated with tested microorganisms. Distilled water at a concentration of 50 μ l/disc provided as a negative control. Plates were incubated at 37°C for 24 h and the formation of clear zone around the disc was observed.

3. RESULTS AND DISCUSSION

The essential oils, having antibacterial effect, against tested bacteria were shown in Table 2. The water (control) was ineffective. Essential oils of thyme (*Thymus vulgaris* L.) and peppermint (*Mentha piperita*) were more effective than others, and in contrast, fennel and cumin essential oils were ineffective against two tested bacteria, *Proteus vulgaris* and *Pseudomonas aeruginosa*. The results showed that the essential oil of peppermint was more effective than thyme essential oil in controlling *Klebsiella pneumoniae*, but for other pathogens the essential oil of thyme was more effective than other essential oils. The investigation of Valero and Salmeron (2003) showed that the essential oil of cinnamon, oregano and thyme were most effective on controlling *B. cereus* [15]. Essawi and Sour reported that *Salvia officinalis*, *Teucrium polium*, *Majorana syriaca*, *Thymus origanium*, *Thymus vulgaris*, *Commiphora opobalsamum*, *Foeniculum vulgare*, and *Rosmarinus officinalis* exhibited an antibacterial effect against both gram-positive and gram-negative bacteria [21]. It has been indicated that the antibacterial activity of some medicinal and aromatic plants are due to different chemical agents in the extracts, including essential oils (especially thymol), flavonoids and triterpenoids and other compounds of phenolic nature or free hydroxyl group, which are classified as active antimicrobial compounds [21-22]. *Escherichia coli*, in comparison other pathogenic bacteria, is more influenced by the essential oils, but *Pseudomonas*

aeruginosa is less influenced by the essential oils. Camporese *et al.* reported that various concentrations of the plants extracts showed activity to some extent against *Escherichia coli* and *Pseudomonas aeruginosa*, while *Aristolochia trilobata* leaves and bark *Syngonium podophyllum* leaves and bark were active also against *Staphylococcus aureus* as well [23]. A recent study demonstrated an antibacterial activity of rosmarinic acid against *Escherichia coli* and *Staphylococcus aureus* [24]. Sagdic *et al.* reported that between extracts of seven spices, thyme and oregano showed higher activity than others on the growth of *Escherichia coli* 0157:H7 strain [14]. Natural substances that extracted from plants (especially, medicinal, aromatic and spice plants) have applications in controlling pathogens in foods [25-26]. Since the medicinal plants studied appear to have a broad anti microbial activity spectrum, they could be useful in antiseptic and disinfectant formulations as well as in chemotherapy [27].

Table 1 Essential oils of plants used in this experiment

Name	Scientific name	Family	Part used for essential oil extraction
Thyme	<i>Thymus vulgaris</i> L.	Lamiaceae	Leaves + Stem
Peppermint	<i>Mentha piperita</i>	Lamiaceae	Leaves
Fennel	<i>Foeniculum vulgar MILL</i>	Apiaceae	Fruits (Seeds)
Caraway	<i>Carum carvi</i> L,	Apiaceae	Fruits (Seeds)

Table 2 Screening for antibacterial activity of essential oils.

Plant essential oil	Diameter of the inhibition zone (mm)			
	<i>Escherichia coli</i>	<i>Proteus vulgaris</i>	<i>Pseudomonas aeruginosa</i>	<i>Klebsiella pneumoniae</i>
Thyme	20	15	13	14
Mint	16	14	12	15
Fennel	8	-	-	6
Cumin	10	-	-	12

4. CONCLUSION

The essential oils of thyme (*Thymus vulgaris* L.) showed the broadest spectrum of action against *Escherichia coli*. The results that reported in this study showed that *Escherichia coli* is more sensitive to essential oils tested, and essential oils of fennel and cumin had no effect on *Pseudomonas aeruginosa* and *Proteus vulgaris*. The obtained results demonstrate that these herbal drugs could represent a new source of antimicrobial agents with less expensive than the imported drugs [23].

REFERENCES

- [1] L.A. Shelef, Antimicrobial Effects of Spices, *Journal of Food Safety*, 6, 1983, 29-44.
- [2] L.L. Zaika, Spices and Herbs: Their Antimicrobial Activity and Its Determination, *J. Food Safety*, 9, 1988, 97-118.

- [3] L.R. Beuchat, and D.A. Golden, Antimicrobial Occurring Naturally in Foods, *Food Technology*, 43, 1989, 134-142.
- [4] E.W.T. Ting, and K.E. Deibel, Sensitivity of *Listeria monocytogenes* to Spices at Two Temperatures, *Journal of Food Safety*, 12, 1992, 129-137.
- [5] B.J. Juven, J. Kanner, F. Schved, and H. Weisslowicz, Factors that Interact with the Antibacterial Action of Thyme Essential Oil and Its Active Constituents, *Journal of Applied Bacteriology*, 76, 1994, 626-631.
- [6] M.M. Cowan, Plant Products as Antimicrobial Agents, *Clinical Microbiology Reviews*, 12, 1999, 564-582.
- [7] C.C. Tassou, E.H. Drosinos, and G.J.E. Nychas, Effects of Essential Oil from Mint (*Mentha piperita*) on *Salmonella enteritidis* and *Listeria monocytogenes* in Model Food Systems at 4 °C and 10 °C, *Journal of Applied Bacteriology*, 78, 1995, 593-600.
- [8] H.W. Chang, Antimicrobial Effect of Spices and Vegetables, *Food Industries (ROC)*, 27, 1995, 53-61.
- [9] A. Sivropoulo, E. Papanikolaou, C. Nikolaou, S. Kokkini, T. Lanaras, and M. Arsenakis, Antimicrobial and Cytotoxic Activities of *Origanum* Essential Oils, *Journal of Agricultural and Food Chemistry*, 44, 1996, 1202-1205.
- [10] A. Chaibi, L.H. Ababouch, K. Bllasri, S. Boucetta, and F.F. Busta, Inhibition of Germination and Vegetative Growth of *Bacillus cereus* T and *Clostridium botulinum* 62A Spores by Essential Oils, *Food Microbiology*, 14, 1997, 161-174.
- [11] J. Wan, A. Wilcock, and M.J. Coventry, The Effect of Essential Oils of Basil on the Growth of *Aeromonas hydrophila* and *Pseudomonas fluorescens*, *Journal of Applied Microbiology*, 84, 1998, 152-158.
- [12] K.J. Lachowicz, G.P. Jones, D.R. Briggs, F.E. Bienvenu, J. Wan, A. Wilcock, and M.J. Coventry, The Synergistic Preservative Effects of the Essential Oils of Sweet Basil (*Ocimum basilicum* L.) Against Acid Tolerant Food Microflora, *Letters in Applied Microbiology*, 26, 1998, 209-214.
- [13] A. Sokmen, B.M. Jones, and M. Erturk, The In vitro Antibacterial Activity of Turkish Medicinal Plants, *Journal of Ethnopharmacology*, 67, 1999, 79-86.
- [14] O. Sagdic, A. Kuscu, M. Ozcan, and S. Ozcelik, Effects of Turkish Spice Extracts at Various Concentrations on the Growth of *Escherichia coli* 0157:H7, *Food Microbiology*, 19, 2002, 473-480.
- [15] M. Valero, and M.C. Salmeron, Antibacterial Activity of 11 Essential Oils Against *Bacillus cereus* in Tyndallized Carrot Broth, *International Journal of Food Microbiology*, 85, 2003, 73-81.
- [16] S. Cosentino, C.I.G. Tuberoso, B. Pisano, M. Satta, V. Mascia, E. Arzedi, and F. Palmas, In vitro Antimicrobial Activity and Chemical Composition of Sardinian *Thymus* Essential Oils, *Lett. Appl. Microbiol.*, 29, 1999, 130-135.
- [17] M. Elgayyar, F.A. Draughon, D.A. Golden, and J.R. Mount, Antimicrobial Activity of Essential Oils from Plants Against Selected Pathogenic and Saprophytic Microorganisms, *J. Food Prot.*, 64, 2001, 1019-1024.
- [18] A. Smith-Palmer, J. Stewart, and L. Fyfe, Antimicrobial Properties of Plant Essential Oils and Essences Against Important Foodborne Pathogens, *Lett. Appl. Microbiol.*, 26, 1998, 118-122.
- [19] E. O'Gara, D.J. Hill, and D.J. Maslin, Activities of Garlic Oil, Garlic Powder, and their Diallyl Constituents Against *Helicobacter Pylori*, *Appl. Environ. Microbiol.*, 66, 2000, 2269-2273.
- [20] A.L. Barry, and C. Thornsberry, Susceptibility Tests Disc Diffusion Test Procedures. In: E.H. Lennette Balows, W.J. Hauster, H.J. Shadomy (Eds.), *Manual of Clinical Microbiology*. American Society for Microbiology, Washington, DC, 1985, 978-987.
- [21] T. Essawi, and M. Srour, Screening of Some Palestinian Medicinal Plants for Antibacterial Activity, *Journal of Ethnopharmacology*, 70, 2000, 343-349.

- [22] A. Rojas, L. Hernandez, R. Pereda-Miranda, and R. Mata, Screening for Antimicrobial Activity of Crude Drug Extracts and Pure Natural Products from Mexican Medicinal Plants, *Journal of Ethnopharmacology*, 35, 1992, 275-283.
- [23] A. Camporese, M.J. Balick, R. Arvigo, R.G. Esposito, N. Morsellino, F. De Simone, and A. Tubaro, Screening of Anti-bacterial Activity of Medicinal Plants from Belize (Central America), *Journal of Ethnopharmacology*, 87, 2003, 103-107.
- [24] R. Li, D. Teng, G. Du, and B. Wang, Isolation of Rosmarinic Acid from Callus of *Perilla frutescens* and Studies of Its Inhibition on the Growth of Bacteria and Fungus, *Weishengwuxue Tongbao*, 27, 2000, 324-327.
- [25] P.M. Davidson, Chemical Preservatives and Natural Antimicrobial Compounds, In: M.P. Doyle, L.R. Beuchat, T.J. Montville (Eds.), *Food Microbiology, Fundamental and Frontiers*, ASM Publications, Washington, DC, 1997, 520-556.
- [26] B.L. Bowles, and V.K. Juneja, Inhibition of Food-borne Bacterial Pathogens by Naturally Occurring Food Additives, *Journal of Food Safety*, 18, 1998, 101-112.
- [27] D.K. Olukoya, N. Idika, and T. Odugbemi, Antimicrobial Activity of Some Medicinal Plants from Nigeria, *Journal of Ethnopharmacology*, 39, 1993, 69-72.