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Research article

New Azo-Schiff Compounds and Metal Complexes Derived from 2-Naphthol Synthesis, Characterization, Spectrophotometric, and Study of Biological Activity

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Abstract

1. Introduction

Azo compounds are one of the most widely used chemical groups of organic compounds, with various applications [1] in advanced organic synthesis to high technology areas such as liquid crystalline displays, ink-jet printers, lasers, and electro optical devices [2]. They are also known to have wide ranges of biological effects such as DNA, RNA inhibition and protein synthesis, carcinogenesis, and nitrogen fixation [3].

 $\frac{1}{\sqrt{2}}$, $\frac{1}{\sqrt{2}}$

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Since azo compounds can form organometallic compounds with transition metal ions, there have been the focus of numerous spectroscopic studies to identify or elucidate structures [4]. Schiff bases are chemical compounds synthesized by condensation of a ketone or aldehyde with a substituted primary amine [5-7], and they were reported by Hugo Schiff in 1864 [8]. Also, Schiff bases are known as the azomethine compounds [9]. The existence of a lone pair of electrons in the azomethine group's sp2 hybridized nitrogen atom orbital is of major chemical and biological significance, according to various studies [10]. Thus, both the azo and Schiff base compounds are known to be antifungal, anti-bacterial [11-14], anti-inflammatory and anti-cancer [15, 16]. In recent years, azo and Schiff base compounds have drawn the attention of researchers to their development and study of application in a variety of fields. For example, Al Zoubi *et al*. [17] prepared new complexes from azo-Schiff base ligand which derived from 4-aminoantypyrine and evaluated as antibacterial activity. Moreover, Alabidi *et al*. [18] developed a new azo molecule obtained from 2-naphthol, which was used to evaluate Zn(II) in pharmaceutical formulations using spectrophotometry. Consequently, this research involved the preparation of azo-Schiff base compounds and their chelate complexes and the study of their biological activities which could be used for the manufacturing of an ointment in order to treat inflammation of wounds and burns caused by pathogenic bacteria.

2. Materials and Methods

The NMR spectra were recorded on a Bruker-NMR 400 MHZ using the solvent DMSO-d⁶. The FT-IR spectra were obtained using a Shimadzu spectrophotometer and a KBr disk. A double beam Shimadzu 1700 UV spectrometer was used to record the UV spectra. TLC was used to check the reaction bath, which was then visualized in iodine vapor. In open capillary tubes, melting points were determined.

2.**1 Chemical synthesis**

2.1.1 Synthesis of azo compound

According to Alabidi *et al*. [19], azo compound (L1) was prepared by dissolving the 4-Amino acetophenone (375 mg, 2.77 mmol) in 20 ml of absolute ethanol and then added to 2 ml of HCl (conc.). Then, stirred for 20 min in an ice bath; after that, an ice-cold solution of sodium nitrite NaNO₂ (10 %) 25 ml was added drop by drop on a period of 35 min. The solution became brown, and was added dropwise to another ice-cold solution of 2- naphthol (400, 2.77 mmol) in 20 ml alkaline ethanol, and constantly stirred at 50°C overnight. Diluted ammonia and hydrochloric acid were used to neutralize the reaction mixture until pH= 7. Afterwards, the resulting product was filtered before being washed in cold distilled water and dried.

Yield: 600 mg 74 %, m.p:141-143 FT-IR (KBr) (cm⁻¹): 3330 (OH), 2931 and 2882 (C-H,aliph), 1652 (C=O, Keto), 1598 (C=C, arom), 1396 (N=N).1 H-NMR, 8.49-8.51 (d,2H , H13+15), 8.23-8.25 (d ,2H, H12+16) , 8.14-8.11 (d,1H, H-4), 8.01-7.98 (d, 1H, H-9, 7.92-7.94 (d, 1H, H-6), 7.78-7.80 (d ,1H , H-3), 7.64-7.66 (t, 1H , H-8), 8.53-8.55 (t ,1H, H-7), 6.83 (s, 1H, OH-arom), 2.41 (s, 3H, Me-Keto). 13C-NMR (100 MHz, DMSO-d6) 196.6 (Keto C-18), 142.5 (C-11), 134.0 (C-14), 132.7 (C-2), 130.1(C-5), 129.6 (C-4), 129.2 (C-10), 128.9 (C-1), 128.1 (C-8), 126.9 (C-7), 125.7 $(C-6+9)$, 122.9 $(C-3)$, 121.8 $(C-13+15)$, 117.1 $(C-12+16)$, 26.6 (Me-18)

2.1.2 Synthesis of azo-Schiff compound (L2) [20, 21]

A solution consisting of L1 (350 mg, 1.206 mmol) in 30 ml of ethyl alcohol was added to another solution (185 mg, 1.206 mmol) of 5-amino salicylic acid, with a drop of acetic acid, and the resultant mixture was heated in reflux for 8-10 h. With chloroform-methanol (4:1) as eluents, the progress of the reaction was observed using TLC. The solid product was collected, filtered, collected, and recrystallized from ethanol to yield the desired product after cooling.

Yield: 550 mg 81 %, yellow powder, m.p:192-194 R.f = 0.35 FT-IR (KBr) (cm⁻¹): 3425 (OH), 2927 and 2783 (C-H aliph), 1660 (C=O, Carboxylic), 1645 (C=N), 1598 (C=C, arom), 1452 (N=N). ¹ H-NMR, 15.88 (br,1H, COOH), 8.42-8.44 (d ,2H , H13+15), 8.17-8.19 (d, 2H, H12+16), 8.05-8.07 (d, 1H, H-4), 7.92-7.94 (d, 1H, H-20), 7.87-7.87 (d, 1H, H-3), 6.73 (s, 1H, OH, C22 arom), 7.58-7.60 (t, 1H, H-8), 7.47-7.49 (t, 1H, H-7), 7.18 (s, 1H, H-24), 6.87-6.89 (d, 1H, H-6), 6.75-6.77 (d, 1H, H-6), 6.67-6.69 (d, 1H, H-21), 6.55 (s, 1H, OH-arom), 2.38 (s, 3H, Me-Keto). 13C-NMR (100 MHz, DMSO-d6) 171.6 (COOH), 154.6 (C=N, C-17), 146.5 (C-23), 142.4 (C-22+11), 138.7 (C-19), 134.0 (C-10), 132.6 (C-2) , 131.5 (C-14), 130.5 (C-21), 130.2 (C-5), 129.6 (C-20+4), 129.2 (C-10), 128.9 (C-1), 128.1 (C-8), 126.9 (C-7), 125.7 (C6+9), 123.4 (C13+15), 122.9 (C-3), 121.8 (C12+16), 26.6 (Me-18).

2.1.3 Synthesis of metal complexes [22]

The metal complexes were obtained by adding 2 mmol of azo-Schiff ligand in 30 ml ethyl alcohol and added it to metal salts drop by drop of cobalt(II), nickel(II), and copper(II) by a stoichiometric amount of (2:1) (ligand: metal). The solution was added to 25 ml of distilled water. Under reflux, the resultant mixture was stirred for 2 h. The products of complexes were collected, washed, purified, and left overnight at 50ºC for dryness. The general equation of synthesized metal complexes is shown below.

$$
MCl_2 + 2HL \rightarrow [ML_2 (H_2O)_2] + 2HCl
$$

 Cu^{+2} complex: FT-IR (KBr) (cm-1): 3446 (OH), 2926 and 2852 (C-H, aliph), 1681 (C=O, carboxylic), 1639 (C=N), 1558 (C=C, arom), 1413 (N=N).

 Ni^{+2} complex: FT-IR (KBr) (cm-1): 3448 (OH), 2924 and 2852 (C-H, aliph), 1678 (C=O, carboxylic), 1595 (C=N), 1498 (C=C, arom), 1440 (N=N).

 Co^{+2} complex: FT-IR (KBr) (cm-1): 3446 (OH), 2931 (C-H, aliph), 1680 (C=O, carboxylic), 1579 (C=N), 1508 (C=C, arom), 1417 (N=N).

2.2. Prepared acid-base solutions [23]

To evaluate the protonation and ionization constants, as well as the effect of pH on the absorption spectra of azo-Schiff compound, multiple pH values of buffer solutions (acetate and universal) were prepared $(1 < -12)$, and concentrations of ligand are (1×10^{-3}) M. The absorbance of these solutions was measured at the wavelength of 300-450 nm as a blank of buffer solution. The constants were determined using the half-length approach.

2.3. Antibacterial activity test

Agar well diffusion has demonstrated the antibacterial activity of ligand, Co, Ni, and Cu complexes [24]. Fifty mg/ml and 200 mg/ml were made from each compound [25]. Two multi-drug resistance pathogenic bacteria were selected: *Staphylococcus aureus* (Gram-positive bacteria) and *Escherichia* *coli* (Gram-negative bacteria) were provided from the Department of Microbiology, Faculty of Science, University of Kufa. Three wells made in Muller-Hinton agar surface by cock borer (10 mm) swabbed with two pathogenic bacteria according to 0.5 McFarland turbidity [26]. Each well received 40 μ L of any dilution, which was incubated at 37°C for 24 h after being left at 20°C for two h. In all tests, the diameter of the inhibition zone was measured in mm done in three replicates [27, 28]. According to SPSS V.10 windows software, a P-value greater than or equal to 0.05 was considered to be significant when compared to the diameters of inhibition zones [29, 30].

3. Results and Discussion

3.1 Chemistry synthesis

To synthesize the azo compound, firstly prepared diazonium salt of 4-aminoacetophenone and coupled it with 2-naphthol stirred in an alkaline solution according to Figure 1.

Figure 1. Synthesis of azo compound

The chemical structure was elucidated by spectral methods. The number of signals in the 1 ¹H-NMR spectra of azo compound (L1) was observed due to the 10 protons back to the three aromatic rings were observed in the δ ~7.53-8.51 ppm range. Some of these doublet signals appeared at chemical shift ($\delta \sim 8.49 - 8.51$ ppm) for H₁₃₊₁₅ due to the withdrawing induced by the keto group. The other triplet signals appeared at the field (δ ∼8.01-7.98, 7.92-7.94 and 7.78-7.80 ppm) due to protons H₉, H₆, and H₃, respectively while the singlet signal of hydroxyl group showed at ($d \sim 6.83$) ppm). At a lower field, Me-keto appeared at $(d \sim 2.41$ ppm), as shown in Figures 2 and 3.

In the ¹³C-NMR spectra, the signal of the carbonyl keto (C_{-17}) appeared at 196 ppm, while the signals back to 16 aromatic carbon atoms were observed at a range of 117-142 ppm. At this time, the signal of the methyl group was shown at 26 ppm (Figures 4 and 5).

Azo-Schiff compound (L_2) was prepared via condensation of 5-aminosalsylic acid with azo derivative (L1) in an acidic solution, according to Figure 6.

Figure 2. Azo compound (L_1) , ¹H-NMR spectra

Figure 3. Azo compound (L_1) , expansion ¹H-NMR spectra

Figure 4. Azo compound, ¹³C-NMR spectra **Figure 5.** Azo compound, expansion ¹³C-NMR spectra

Figure 6. Synthesis of azo- Schiff compound

¹H- NMR spectra showed a new broad signal at the downfield ($\delta \sim 15.88$ ppm) back to the proton of the carboxyl group; the bands of H_{13,15} and H_{12,16} appeared at a field less than the field of derivative (L1) at shift (δ ∼8.42-8.44, and 8.17-8.19 ppm), respectively. Because of a change in the neighboring group from C=O to C=N, the new other singlet group appeared at the field ($\delta \sim 7.18$) ppm) due to H-24 of benzoic acid derivative, as shown in Figures 7 and 8.

The 13C-NMR spectra showed new signals at the downfield of the spectrum at shifts 171and 154 ppm due to the carboxyl (C_{25}) and azomethane (C_{17}) groups, respectively. Meanwhile, the signals at the range of $\delta \sim 115-146$ ppm assigned to 22 carbon atoms of four aromatic rings, also the signal of methyl group was showed at $\delta \sim 26$ ppm, as seen in Figures 9 and 10.

446 172
076 $H - 4/d$ $H-24(s)$ $H-21(d)$ $H3/d'$ $H-9(d)$ $H-6(d)$ OHC-22 H-8(t) $H13+15(d)$ $H12+16d$ $H - 7(t)$ $\begin{array}{c} 3.29 \\ 2.22 \\ 4.34 \end{array}$ $\frac{1}{1.8}$ ic's $\frac{1}{2}$ 3.0% 3.5 2.8 $3,80$ $\frac{1}{2}$ 8.50 7.50

Figure 7. Azo-Schiff compound, ¹H-NMR spectra

Figure 8. Azo-Schiff compound, expansion ¹H-NMR spectra

Figure 9. Azo-Schiff compound, ¹³C-NMR spectra

Figure 10. Azo-Schiff compound, expansion ¹³C-NMR spectra

3.2 FT-IR Spectra

FT-IR of complexes (Figures 11-13) appeared very different from the ligand shown in Figure 14. The peak of hydroxyl and carbonyl groups shifted to the higher wave number, which can be due to hydrogen bonding [31]. The wave number of azo groups shifted to the lower as a result of the coordinate. Therefore, the chemical structure of complexes can be found in Figure 15 via ligand: metals (2:1), so the hybridization of complexes is sp^3d^2 , and the shape is octahedral [32].

3.3 Absorption spectra of azo-Schiff compound

The electronic absorption of ligand clearly shows three bands (Figure16) at different wavelengths. The n- π^* transition was assigned to the first and second bands, which appeared at 206 and 320 nm, respectively. The other band observed at 486 nm, assigned π -π* transition, represents the λ _{max} of ligand [33]. This band displayed a red shift during metal ions coordination, as shown in Figures17- 19.

Figure 13. FT-IR of Cu(II) complex **Figure 14.** FT-IR of azo- Schiff compound

Figure 15. Chemical structure of complexes

Figure 16. Absorption spectra of azo-Schiff **Figure 17.** Absorption spectra of Co(II)

Figure 18. Absorption spectra of Cu(II) complex

 500.0

400.0

 600.0

700.0

 800

Figure 19. Absorption spectra of Ni(II) complex

3.4 Absorption spectra of azo-Schiff ligand in buffer solution

A series of buffer solutions were prepared at various pH to study the effect of pH on the azo-Schiff compound. The absorbance of concentration with the ligand is $(1 \times 10^{-3} - 2 \times 10^{-3})$ at the wavelength of 300-450 nm, which represents graphically in Figure 20. The carboxyl group appeared at pH 7-8, while the hydroxyl group of naphthol appeared at pH 8-9. However, the hydroxyl group of benzoic acid derivative appeared at pH 10-11. In addition, the protonated forms appear at low pH values (1- 4) (Figure 21). The results shown in Table 1 suggested the effect of pH on the azo-Schiff compound as shown in Figure 22.

Figure 20. Acid-base properties of azo- Schiff compound

Figure 21. Absorption spectra of azo-Schiff compound at varying pH values

Table 1. The protonation (pk_p) and ionization (pk_a) constants of the Azo-Schiff compound

Figure 22. Absorbance-pH curves of azo-Schiff compound

The protonation and ionization constants [23] were estimated using the half-height approach to determine pka values from the built absorbance–pH curves in Figure 22. The relationship: $pK_a = pH$ at $(A_1/2)$ when $(A_1/2) = (A_1+A_{min})/2$. Al means limited absorbance, and Amin means minimum absorbance.

3.5 Antibacterial activity

Our findings demonstrated that all derivative compounds have no antibacterial effect on *Escherichia coli*. While only Cu and Co have an antibacterial effect against *Staphylococcus aureus* at both concentrations (Figures 23 and 24). According to statistical analysis, the results indicate that the diameter of inhibition zones of Cu 50 mg/ml (12.967 \pm 0.29) and 100 mg/ml (13.267 \pm 0.39) had no statistically significant differences (P-value 0.2863) against *S. aureus* (Figure 25). In addition, there was no significant difference (P-value 0.1336) between the diameter of inhibition zones of Co 50 mg/ml (10.83±0.33) and 100 mg/ml (11.60±0.49) against *S. aureus* Figure 26. On the other hand, a significant increase (P-value 0.0042**) has been indicated between the diameter of inhibition zones in Cu 50 mg/ml as compared with Co 50 mg/ml against *S. aureus* (Figure 27). In addition,

Figure 23. Antibacterial activity of Cu 100 mg/ml against *S. aureus*

Figure 24. Diameters of inhibition zones of Co against *S. aureus*

 11.60 ± 0.49

Diameters of inhibition zone (mm) Diameters of inhibition zone (mm) 10.83 ± 0.33 **10 5 0 50 100** Concentration mg/ml

15 P-value 0.1336

Figure 25. Diameters of inhibition zones **Figure 26.** Diameters of inhibition zones of Cu against *S. aureus* of Co against *S. aureus*

Figure 27. Comparison between diameters of inhibition zones of Cu and Co against *S. aureus*

when Cu 100 mg/ml was compared to Co 100 mg/ml against *S. aureus*, a significant increase (Pvalue 0.0287*) in the diameter of inhibitory zones was observed.

4. Conclusions

Three main complexes were synthesized in high yields, and the products were analyzed using several spectrum methods. Based on UV-Vis, FTIR, ¹HNMR, and ¹³CNMR spectroscopy, a structure for complexes was proposed, showing that the N and O atoms in the azo-Schiff ligand bonded with metal ions. The (L_2) and its complexes were studied for antibacterial activity against *E. coli* and *S. aureus*. The antimicrobial screening revealed that none of the three complexes have the antibacterial activity against *E. coli* whereas Cu(II) and Co(II) complexes have the antibacterial activity against *S. aureus.*

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