# **Research article**

# Effect of Bleaching Containing Polydopamine and Chitosan-Modified TiO<sub>2</sub> on the Level of Brightness and Microhardness of Teeth

Lydia Rohmawati<sup>1</sup>\*, Fitria Tahta Alfina<sup>1</sup>, Devi Ragita Putri Pratiwi<sup>1</sup>, Angela Arin Pratama<sup>1</sup>, Fariz Irkham Muadhif<sup>1</sup>, Woro Setyarsih<sup>1</sup>, Munasir<sup>1</sup>, Prima Nerito<sup>2</sup> and Darminto<sup>3</sup>

<sup>1</sup>Department of Physics, Faculty of Mathematics and Natural Sciences, Universitas Negeri Surabaya, Ketintang, Gayungan, Surabaya 60231, Indonesia <sup>2</sup>Department of Dentistry, Faculty of Dentistry, Universitas Muhammadiyah Surabaya, Sutorejo, Mulyorejo, Surabaya 60113, Indonesia <sup>3</sup>Department of Physics, Faculty of Science and Data Analytics, Institut Teknologi Sepuluh Nopember (ITS), Keputih, Sukolilo, Surabaya 60111, Indonesia

Curr. Appl. Sci. Technol. 2024, Vol. 24 (No. 6), e0260818; https://doi.org/10.55003/cast.2024.260818

Received: 6 October 2023, Revised: 8 March 2024, Accepted: 24 May 2024, Published: 16 August 2024

## Abstract

Teeth bleaching techniques generally use high concentrations of Keywords bleaching agents such as H<sub>2</sub>O<sub>2</sub>, which can harm dental health. Therefore, an alternative method is needed to minimize the use of H<sub>2</sub>O<sub>2</sub>. The aim of bleaching; this study was to determine the characteristics and effectiveness of a teethwhitening gel made from polydopamine and chitosan-modified TiO<sub>2</sub>. The microhardness; research phase began with the extraction of TiO<sub>2</sub> from Tulungagung sand brightness level using the leaching method, and then the TiO<sub>2</sub> was modified with polydopamine and chitosan. XRD, FTIR, and TEM were used to characterize the fabrication results. The results of XRD analysis showed that the diffraction peaks of polydopamine and chitosan-modified TiO<sub>2</sub> had the characteristics of anatase phase TiO2. Functional groups of polydopamine and chitosan-modified TiO<sub>2</sub> were identified from the results of FTIR analysis. The TEM image showed the spherical shape with a core-shell structure, where the TiO<sub>2</sub> particles were covered with polydopamine and chitosan. The addition of H2O2 at 3% to the polydopamine and chitosan-modified TiO<sub>2</sub> gel transformed it into a tooth whitening agent. After that, the teeth without soaking and those soaked in cola were bleached with the whitening gel using visible light irradiation three times for 15 min each time. The bleaching results showed that the 0.25-g polydopamine-modified TiO<sub>2</sub> formula whitening gel effectively whitened teeth without causing a significant change in the microhardness value of the tooth surfaces, even at low concentrations of H<sub>2</sub>O<sub>2</sub>.

E-mail: lydiarohmawati@unesa.ac.id

<sup>\*</sup>Corresponding author: Tel.: (+62) 81235406463

# 1. Introduction

Dental treatment with bleaching procedures is a conservative method that has been well-accepted by patients and has been confirmed to be the safest and most effective compared to dental procedures either by direct or indirect restoration of tooth color [1, 2]. Teeth bleaching can be done at home or in an orthodontic clinic [3, 4]. At the office, the bleaching chemicals are used in high concentrations to shorten treatment time. On the other hand, home bleaching at a low concentration of 10% is used. Dental bleaching at home may require treatment for extended periods, for example, 4-8 h a day for 14 days or more [5]. Although the home whitening treatment is the most commonly used method, some patients prefer teeth whitening to be performed in a clinic, where the results are quicker [3]. The active component of most whitening products is hydrogen peroxide (H<sub>2</sub>O<sub>2</sub>). Although the use of H<sub>2</sub>O<sub>2</sub> provides optimal results on both vital and non-vital teeth, there are some concerns regarding extensive whitening techniques. This is because H<sub>2</sub>O<sub>2</sub> action on tooth surfaces [6] is triggered by free radicals [7], which can cause tooth oversensitivity and changes in enamel structure, such as changes in hardness and micro-roughness of the teeth [7], as well as injuries to the gums [8, 9].

Therefore, to minimize side effects due to higher concentrations of  $H_2O_2$  (30-35%), alternative materials that have the potential to whiten teeth, such as TiO<sub>2</sub> nanoparticles, are needed [10]. TiO<sub>2</sub> was previously extracted from Tulungagung ilmenite sand, which was known to contain TiO<sub>2</sub> at 12.2% [11]. The advantage of the use of the leaching method in the synthesis of TiO<sub>2</sub> from Tulungagung ilmenite sand, as described by Rohmawati *et al.* [12], was a simple method that did not have a high temperature roasting step. Magnetic and non-magnetic materials from ilmenite sand were separated using a magnetic bar. The result of the separation was a magnetic material that was then dissolved with sulfuric acid. The extraction products from ilmenite sand were 100% anatase TiO<sub>2</sub> without any impurities. TiO<sub>2</sub> phase could be obtained from ilmenite sand by the use of hydrothermal method with NaOH solvent [13] and the caustic fusion method with hydrochloric acid solvent [14]. However, impurities still remained after the use of both methods. Apart from the simplicity of the hydrothermal leaching method used for synthesizing TiO<sub>2</sub> anatase mentioned above, there is also an abundance of sand that has yet to be utilized optimally because so far it has only been used as a building material in the area.

Titanium dioxide (TiO<sub>2</sub>) is a nontoxic biocompatible compound that possesses antimicrobial properties [15, 16]. Over the last few decades, researchers have focused on TiO<sub>2</sub> because of its nature as a photocatalyst and because of its many potential applications in the biomedical field [17]. TiO<sub>2</sub> nanoparticles affect the efficiency of whitening agents without reducing the hardness of the tooth enamel surface [18]. Bleaching teeth with visible light irradiation using a concentration of 35% H<sub>2</sub>O<sub>2</sub> is less effective in whitening teeth when compared to the use of a 35% H<sub>2</sub>O<sub>2</sub> formula and TiO<sub>2</sub> with UV light irradiation [19] because when TiO<sub>2</sub> is exposed to UV light, it actively forms free radicals [16]. The photocatalyst properties of TiO<sub>2</sub> nanoparticles can increase with prolonged use of UV. However, the clinical use of UV light for teeth whitening has undesirable effects [20]. Long-term exposure to UV radiation results in toxic irritation that leads to cell damage, immune suppression, skin cancer, and photoaging [21], as well as causing problems in soft tissues, including the oral mucosa [22, 23].

Zhang *et al.* [24] reported that using visible light for 30 min was a solution to replace UV light with an additional 30% H<sub>2</sub>O<sub>2</sub>. The research results provided a white effect, but damage occurred to tooth enamel due to high levels of H<sub>2</sub>O<sub>2</sub>. Likewise, a concentration of 6% H<sub>2</sub>O<sub>2</sub> in a TiO<sub>2</sub> solution for 45 min in visible light showed a white effect, but the teeth experience overbleaching, eroding the enamel [8]. Suemori *et al.* [25] carried out tooth whitening from TiO<sub>2</sub> with the addition of 3.5% H<sub>2</sub>O<sub>2</sub>, each activated by 405 nm diode laser light and halogen lamp for 15 min, showing a level of tooth brightness without damaging the enamel. Sun *et al.* [26] described that the

photocatalytic activity of TiO<sub>2</sub> anatase in visible light could be improved by modifying it with polydopamine.

Polydopamine (PDA) has been used in various applications because it is considered a natural biopolymer [27]. PDA is a polymer that can be applied in several materials, such as metals, metal oxides, non-metal oxides, silica, ceramics, polymers, and other nanomaterials [28]. PDA has optical properties related to its absorption of ultraviolet (UV) light and visible light [27], and can be used with TiO<sub>2</sub> to improve catalysis in short irradiation process. Zhang *et al.* [24] reported that PDA-modified nano TiO<sub>2</sub> can increase the brightness of the teeth. The results of their study show that when the sample is still in the form of powder, it is less efficient at teeth whitening. Teeth whitening gel with 6% H<sub>2</sub>O<sub>2</sub>, including TiO<sub>2</sub> and chitosan, provided compelling whitening without negative effects on the roughness or hardness of the tooth surface [15]. Chitosan is an alternative remineralizing teeth agent that is of low cost [29] and does not irritate the gums [30]. Chitosan is a natural polysaccharide, non-toxic, biocompatible, and biodegradable.

Chitosan is widely used in biomedical, food, cosmetic, and pharmaceutical applications because of its film and gel-forming capabilities. It is also bioactivity and used as bioadhesive. Moreover, it possesses remineralizing and antibacterial characteristics [31-33]. Chitosan was used in experimental gel whitening agents as an alternative to synthetic polymers. It functions as a thickener, carrier, bioadhesive, demineralizing, antioxidant, and antimicrobial [15]. In addition, chitosan-modified TiO<sub>2</sub> displays a combination of the photocatalytic properties of nano TiO<sub>2</sub> and the adsorption properties of chitosan, making it very appropriate and adequate for bleaching care [34]. Li *et al.* [35] described that adding 0.05-0.2% chitosan expanded the brightness of the Kraft pulp. Based on information from several researchers above, this research fabricated PDA and chitosan-modified TiO<sub>2</sub> with 3% H<sub>2</sub>O<sub>2</sub> as a tooth whitener, which can later be developed into a home dental care product. So far, the use of PDA and chitosan-modified TiO<sub>2</sub> with 3% H<sub>2</sub>O<sub>2</sub> has never been reported. Thus, in this research, the influence of PDA and chitosan-modified TiO<sub>2</sub> with 3% H<sub>2</sub>O<sub>2</sub> on the brightness level and microhardness of the teeth was determined and the results of this teeth whitening product are expected to be effective in brightening teeth and does not damaging the teeth.

# 2. Materials and Methods

#### **2.1 Materials**

The materials used in the manufacture of PDA and chitosan-modified  $TiO_2$  were Tulungagung Ilmenite sand,  $H_2SO_4$  (Sigma Aldrich 99%), dopamine hydrochloride (Sigma Aldrich 99%), hexamethylenetetramine (Sigma Aldrich 99%), NaOH (Merck), CH<sub>3</sub>COOH (Merck), ethanol (Merck), distilled water, cola, and molar teeth. Equipment for the experimental process included beakers, a digital Ohaus balance, measuring cups, laboratory spatula, mortar and pestle, 200 mesh sieves, a vacuum pump, a glass filtration unit, hot plates, Buchner funnels, a drying oven, and a furnace.

#### 2.2 Synthesis of anatase TiO<sub>2</sub>

TiO<sub>2</sub> manufacture was based on the method of Rohmawati *et al.* [12] by the use of the hydrothermal leaching method. This involved pulverizing ilmenite sand using a mortar and pestle and then sifting the sand to a size of 200 mesh. After that, the sand was dissolved in 8 M H<sub>2</sub>SO<sub>4</sub> and stirred using a magnetic stirrer at 700 rpm at 120°C. The solution formed a slurry and the TiOSO<sub>4</sub> filtrate and FeSO<sub>4</sub> precipitate were then separated using a vacuum pump. The filtrate from the separation was added to distilled water and heated at 300°C for 12 h. The precipitate formed by the heating process

was washed with distilled water several times until its pH was 7. The solution was then filtered, and the residue was calcinated at 600°C for 2 h. A dry powder was then obtained. The stages of the anatase TiO<sub>2</sub> synthesis process are illustrated in Figure 1.



Figure 1. Preparation process for the synthesis of TiO<sub>2</sub> from Tulungagung ilmenite

# 2.3 Fabrication of polydopamine and chitosan-modified TiO<sub>2</sub>

A mass of TiO<sub>2</sub> of 0.08 g and a mass of dopamine of 0.04 g were mixed with distilled water and stirred until a suspension was formed [36]. Then, 0.1 g of hexamethylenetetramine (HMTA) was added to the solution using the sonication method. The solution was then dried at 90°C for 3 h in a drying oven; then, the solution was centrifuged at 4000 rpm to produce precipitates. Next, the precipitate was washed with distilled water and ethanol to acquire a wet precipitate. After that, it was dried in the drying oven at 80°C for 30 min and PDA-modified TiO<sub>2</sub> was obtained (Figure 2).

In the next stage, 0.2 g of chitosan was first dissolved in 7 mL of acetic acid and 25 mL of distilled water. Then, 0.05 g of PDA-modified TiO<sub>2</sub> was added. After that, the mixture was stirred at 60°C with a speed of 500 rpm and 0.1 M NaOH was added until the pH solution was 7, and the brownish-colored gel obtained. In the next stage, 3% H<sub>2</sub>O<sub>2</sub> was added to the gel and stirred until homogeneous. The above steps were repeated for PDA-modified TiO<sub>2</sub> with a mass of 0.25 g. The detailed stages of fabrication of PDA and chitosan-modified TiO<sub>2</sub> can be seen in Figure 3.

#### 2.4 Characterization

TiO<sub>2</sub>, PDA-modified TiO<sub>2</sub>, and PDA and chitosan-modified TiO<sub>2</sub> samples from the synthesis were characterized by XRD (X-ray diffraction) to determine the diffraction patterns of the anatase TiO<sub>2</sub> phase and changes in the TiO<sub>2</sub> crystal structure due to modification with PDA and chitosan. The XRD device used was a Philips X'Pert MPD system with a Cu anode radiation source of 40 kV, 30 mA, and a wavelength of CuK $\alpha$  of 1.54056 Å at an angle of 2 theta 10-90°. In addition, FTIR (Fourier transform infrared spectrophotometry) was also performed to determine the functional groups of the fabricated samples. FTIR results were recorded on a Shimadzu-type IR Prestige 21 instrument with a 4000-500 cm<sup>-1</sup> wavenumber range. TEM (transmission electron microscopy) (Tecnai G2 20S-Twin Function type) was performed to determine the morphology and the particle size of PDA and chitosan-modified TiO<sub>2</sub> samples and ImageJ software was used.



Figure 2. Preparation of TiO<sub>2</sub> by modified polydopamine



Figure 3. Fabrication of polydopamine and chitosan-modified TiO<sub>2</sub>

Teeth with and without cola immersion underwent bleaching. The bleaching result was then measured at the brightness level using a UV-vis spectrophotometer (Ultraviolet-visible spectroscopic PC-2401). The data obtained from spectrophotometer were calculated for color parameters at distances L\*, a\*, b\*, based on the CIELAB system created in 1978 by the Commission International de I'Eclairage (CIE). The CIELAB system explains color perception in three dimensions. The L\* value denotes the brightness level of the teeth (light or dark), a\* in color analysis represents the red-green level, and b\* defines the yellow-blue level. The level of tooth microhardness after bleaching was determined by a Vickers hardness test (HV) on a Mitutoyo HM 200. Before the microhardness test, the teeth were cut in half from the tooth crown and then coated with resin as an adhesive during the test. The hardness test was conducted for 20 s with a loading force of 0.1 N to appraise superficial <5  $\mu$ m subsurface enamel [37]. The level of surface abrasion of tooth enamel was determined using a scanning electron microscope (Zeiss EVO MA10) with a voltage range of 0.2 to 30 kV.

# 3. Results and Discussion

# 3.1 X-ray diffraction (XRD)

In this study, XRD characterization was carried out to determine the structure of the main phase in the synthesized sample, and Match! software was used for data analysis. The sample diffraction spectra in Figure 4 shows that each peak indicates the main phase of TiO<sub>2</sub> anatase according to JCPDS (Joint Committee on Powder Diffraction Standards) Number 96-900-8216. The diffraction peaks are 25.28, 37.58, 47.86, 53.59, 54.90, 62.33, 68.40, 74.86, 82.28 with the Miller index (101), (004), (020), (015), (121), (024), (116), (125), (224), respectively, which together characterize the anatase phase. The highest diffraction peak in-plane orientation (101) was located at an angle of 25.28°, which was in agreement with the results of Saranya *et al.* [38] and Kalaiarasi and Jose [39], who reported angles of 25.25° (101) and 25.3° (101), respectively. The diffraction peaks for the PDAmodified  $TiO_2$  sample show the characteristics of the anatase  $TiO_2$  phase. The peaks are similar to the standard TiO<sub>2</sub> diffraction pattern peaks, so the addition of PDA had no impact on the TiO<sub>2</sub> crystal structure. Likewise, the diffraction pattern of the PDA and chitosan-modified TiO<sub>2</sub> samples did not show a change in TiO<sub>2</sub> crystallinity; however, the intensity of the diffraction peaks changed due to the presence of H bonds between chitosan and  $TiO_2$  [40]. The chitosan diffraction peaks were not found in the diffraction patterns of PDA and chitosan-modified by TiO<sub>2</sub> samples. Ths indicated that bonds between chitosan, polydopamine, and TiO<sub>2</sub> had formed.



Figure 4. Diffraction patterns of PDA-modified TiO<sub>2</sub> and PDA-chitosan-modified TiO<sub>2</sub> against standard TiO<sub>2</sub>

### **3.2** Fourier transform infrared spectrophotometry (FTIR)

The FTIR characterization was used to identify groups of chemical bonds in the TiO<sub>2</sub> samples, PDAmodified TiO<sub>2</sub> (TiO<sub>2</sub>-PDA), and PDA and chitosan-modified TiO<sub>2</sub> (TiO<sub>2</sub>-PDA-chitosan). The data from the test results in Figure 5 shows the absorption peaks over the wavenumber range 4000-500 cm<sup>-1</sup>. The experimental sample peaks were identified by matching them with reference data, as detailed in Table 1. The TiO<sub>2</sub> samples synthesized from ilmenite sand in the Tulungagung coastal area contained organic compounds that usually occur in seawater organisms, one of which was detected in the FTIR spectrum in Figure 5(a), with absorption peaks at 1180.44 and 1114.86 cm<sup>-1</sup>, propably indicating an alkyl halide compound [41]. At wavenumber 1055.06 cm<sup>-1</sup>, an absorption peak which indicated O-H stretching vibrations was observed. This was probably was due to the presence of absorbed water and hydroxyl groups [42]. The Ti-O stretching and Ti-O-Ti peaks, characteristics of TiO<sub>2</sub> anatase, were found at 935.48, 825.53, and 754.17 cm<sup>-1</sup>.

Figure 5(b) shows the spectrum of the PDA-modified TiO<sub>2</sub> sample where the absorption peaks at 935.48 and 771.53 cm<sup>-1</sup> indicate the functional groups of TiO<sub>2</sub> anatase. The characteristic peaks of PDA are at wave numbers of 1456-1058 cm<sup>-1</sup>. These corresponded to the functional groups of C-C, CH<sub>2</sub> scissoring, C–O symmetry stretching vibration, C-O vibration, and C-H plane bending vibration. The TiO2-PDA-chitosan sample in Figure 5(c) showed absorption peaks at 3642.25 and 3562.52 cm<sup>-1</sup>, showing hydrogen bonds between the –H bonded hydroxyl groups due to the absorption of H<sub>2</sub>O [43]. Group of C=O stretching in the amide I was identified at wavenumbers of 1990.54, 1687.7, 1643.35, and 1614.42 cm<sup>-1</sup>, which were wavenumber characteristics of chitosan [44]. The attributes of PDA were identified at 1091.7 cm<sup>-1</sup> in the C-H functional group. The absorption peaks at 590.22, and 540.07 cm<sup>-1</sup> indicated groups belonging to TiO<sub>2</sub> [45]. Therefore, the results of the FTIR analysis above showed that the functional group of PDA and chitosan-modified TiO<sub>2</sub> samples were identified where each absorption peak shows the characteristics of polydopamine and chitosan-modified TiO<sub>2</sub>.



**Figure 5.** The FTIR transmittance spectra of the samples (a) TiO<sub>2</sub> (b) PDA-modified TiO<sub>2</sub> (c) PDA and chitosan-modified TiO<sub>2</sub>

Sample	Wavenum	ber (cm <sup>-1</sup> )	Functional Group			
	Experiment	Reference				
TiO <sub>2</sub>	1180.44, 1114.86	1104 [41]	Alkyl halide			
PDA-modified TiO <sub>2</sub>	1055.06 935.48, 825.53, 754.17 1456.26	1078 [42] 1000-400 [46] 1450 [47]	O-H stretching vibrations Ti-O stretching, Ti-O-Ti bridging stretching C-C			
	1292.31 1195.87	1288 [48] 1184 [49]	CH <sub>2</sub> scissoring C–O symmetry stretching vibration			
	1122.57 1091.71, 1058.92	1120 [50] 1065 [51]	C-O vibration C-H plane bending vibration			
	935.48, 771.53	1000-400 [46]	Ti-O stretching, Ti-O-Ti bridging stretching			
PDA and chitosan- modified TiO <sub>2</sub>	3642.25, 3562.52	3700-3420 [43]	-H bonded hydroxyl group			
	1990.54, 1687.7, 1643.35, 1614.42	1645 [44]	C=O stretching of amide I			
	1556.55 1149.57	1550 [44] 1153 [52]	N–H bending in amide II Asymmetric stretching C-O-C			
	1091.7 590.22 540.07	1065 [51] 740.67 [44] 609.51, 516.92 [45]	C-H plane bending vibration Ti–O vibration TiO <sub>2</sub> stretching			

Table 1. Functional groups on FTIR wavenumbers

## 3.3 Transmission electron microscopy (TEM)

The morphology and coating forms of PDA and chitosan-modified  $TiO_2$  samples and the particle size of the composite was observed using TEM, as shown in Figure 6. The particle size distribution was determined by Image J software, and the analysis was based on the number of 100 grains.

Figure 6 shows that the PDA and chitosan-modified TiO<sub>2</sub> samples have a core-shell structure. The PDA and chitosan-modified TiO<sub>2</sub> particles were of spherical shape. These results were in alignment with the research of Zhang *et al.* [24], who synthesized a PDA-modified nano TiO<sub>2</sub> (nano-TiO<sub>2</sub>@PDA) material that had a particle size of 40 nm and particle size thickness of about 2 nm. In our study, the TEM analysis in Figure 7 clearly shows that the shape of the TiO<sub>2</sub> particles covered by polydopamine and chitosan had a particle core size of 70.63 nm, while an organic surface layer (shell) had a surface thickness of 7.62 nm. An increased TiO<sub>2</sub> particle size may be caused by TiO<sub>2</sub> agglomeration occurring with an irregular size distribution [53]. Thus, the PDA and chitosan-modified TiO<sub>2</sub> in our study can be considered a nano material because of its size of  $\leq 100$  nm.



Figure 6. Micrograph of TiO<sub>2</sub>-PDA-chitosan sample



**Figure 7.** Particle size distribution of PDA and chitosan-modified  $TiO_2(a)$  shell size (b) core size

# 3.4 Tooth brightness level using the PC-2401 UV-vis spectrophotometer

The results of the teeth bleaching process using PDA and chitosan-modified  $TiO_2$  formulas with 3%  $H_2O_2$  and containing 0.25 g and 0.05 g, are shown in Table 2. The samples were bleached 3 times for 15 min under visible light irradiation, and applied to the tooth surface. Brightness level of the teeth was determined using the UV spectrophotometer. The value of tooth brightness (L\*) before bleaching was 112.34 for the tooth sample without cola immersion, whereas after bleaching with the formula 0.05-g tooth whitening gel, it became 112.96. Likewise, teeth soaked in cola had a brightness level of 103.67 before bleaching, while after bleaching, the 0.05-g formula whitening gel gave a brightness of 112.48. The results obtained from the 0.25 g of whitening gel formula showed significant brightness with and without immersion treatment in cola, namely 121.77 and 121.25.

When  $TiO_2$ -based tooth whitening gel adheres to colored tooth enamel and is exposed to blue visible light, its  $TiO_2$  becomes photocatalytically active. The photon energy absorbed by  $TiO_2$  causes electrons to be excited into a conduction band, thereby reducing oxygen levels and causing

Sample	Condition	Teeth Whitening	<b>Teeth Color Test Value</b>					
		Gel Content (g)	L*	a*	b*	dE*ab		
Teeth without cola soak	Before bleaching	-	112.34	-0.62	0.06	0.00		
Teeth with cola soak for 7 days	After bleaching Before bleaching	0.05 0.25 -	112.96 121.77 103.67	-1.01 -2.87 -0.94	- 1.23 -4.75 -1.19	1.48 10.82 0.00		
	After	0.05	112.48	-2.78	-10.12	12.67		
	bleaching	0.25	121.25	-4.84	-10.61	20.32		

Tabl	e 2	. Т	he res	sults	of	quanti	tative	test	of	tooth	i brig	ghtness	using	U	V-v	is l	PC	2
------	-----	-----	--------	-------	----	--------	--------	------	----	-------	--------	---------	-------	---	-----	------	----	---

production of superoxide radicals ( $O_2^-$ ), while holes formed in the valence band cause a decrease in hydroxide ions and the production of hydroxyl radicals (OH<sup>-</sup>). These free radicals can modify organic compounds such as chromogens that cause discoloration of teeth through oxidation and degradation processes [10], in which the chromogen molecules are degraded into smaller, transparent, and soluble molecules, removing stains and whitening teeth [54]. Teeth whitener can produce free radicals even though it only uses a concentration of 3% H<sub>2</sub>O<sub>2</sub> [10]. The addition of PDA in this research likely increased the catalytic activity of TiO<sub>2</sub>, especially during the bleaching process using visible light [24]. Moreover, the addition of chitosan can combine the photocatalytic properties of nano TiO<sub>2</sub> and the adsorption properties of chitosan, so it is a suitable and effective additive for whitening treatments [15, 34] and it can even increase the adhesion of the gel onto the tooth surface, thereby stabilizing and even preventing the release of free radicals [35]. In addition, chitosan can also function as a thickener, bioadhesive, and remineralization agent [15]. However, using high catalyst concentrations is not recommended because such concentrations can disrupt the aggregation of catalyst particles, significantly reducing photocatalytic activity [55-57].

The tooth brightness value obtained in this study was greater than that of Suemori *et al.* [25], who used TiO<sub>2</sub> and 3.5% H<sub>2</sub>O<sub>2</sub> and Ozcetin and Surmelioglu [15], who examined a formula with TiO<sub>2</sub>/chitosan/6% HP. In Suemori *et al.* [25], a TiO<sub>2</sub> formula and 3.5% H<sub>2</sub>O<sub>2</sub> were used for bleaching under visible light, as was done in this study. Their obtained data were processed with the Tukey test including the highest brightness of 72.24<sup>cd</sup> after visible light irradiation produced by a 405 nm diode laser, whereas under a halogen lamp (320-1100 nm), the teeth had a brightness of 71.04<sup>cd</sup>. Ozcetin and Surmelioglu [15] used tooth whitening gel under UV light (< 385 nm) to form free radicals with a brightness of 90.5±5.1. However, continuous UV irradiation has an unfavorable effect around the teeth [58]. PDA and chitosan-modified TiO<sub>2</sub> can increase the brightness of the teeth, even though the bleaching process is carried out by irradiating visible light. This is because PDA and chitosan-modified TiO<sub>2</sub> can produce free radicals that degrade the color of teeth, making them appear whiter and brightner [59].

Teeth whitening in this study was repeated thrice every 15 min. For each repetition, the teeth were cleaned and dried, then bleached again for 15 min. The effect of bleaching on tooth brightness was observed for each repetition, both for teeth with and without cola soaking treatment. The length of exposure during the bleaching process can increase the photocatalytic activity of the teeth-whitening gel, producing many free radicals. However, the effect of prolonged exposure can increase the temperature along with the penetration of  $H_2O_2$ , resulting in an increase in pulp temperature and causing the teeth to become sensitive [60].

## 3.5 Microhardness of teeth

Tooth samples that had gone through the bleaching process were subjected to the Micro–Vickers hardness test to determine the hardness of the teeth. The Micro-Vickers test was applied using a loading force of 0.1 N to determine the subsurface hardness of superficial enamel <5  $\mu$ m. microhardness test results data can be seen in Table 3 for teeth before and after the bleaching process, both for teeth without cola immersion and teeth with cola immersion.

Table 3. Microhardness values before and after the bleaching process

Teeth Sample	Condition	Teeth Whitening Gel (g)	Microhardness (HV)
Teeth without	Before bleaching	-	16.67
cola soak	After bleaching	0.05	16.27
		0.25	16.13
Teeth with cola	Before bleaching	-	26.17
soak for 7 days	After bleaching	0.05	25.43
		0.25	25.20

The results of the study presented in Table 3 show the microhardness of the teeth samples without immersion and with cola immersion for 7 days. Before the bleaching process, the teeth without cola immersion were tested for microhardness, and a microhardness value of 16.67HV was obtained. However, after the bleaching process, the microhardness of teeth after being smeared with the 0.05-g whitening gel material was 16.27 HV. Similarly, for 0.25-g whitening gel material, the tooth microhardness obtained was 16.13 HV. In the next test, teeth soaked in cola for 7 days were studied. Teeth were immersed to the roots and the hardness and abrasion of the teeth was evaluated. Based on the hardness test results, teeth soaked in cola before bleaching had a hardness of 26.17 HV. After bleaching and using a composition of 0.05 g and 0.25 g of teeth-whitening gel formulas, microhardness of 25.43 HV and 25.20 HV, respectively, was observed. The tooth hardness test obtained in this study showed low values because the loading force of the test was carried out on the superficial enamel  $< 5 \,\mu\text{m}$ . The use of bleaching agent concentration [61, 62], application time [63], and bleaching agent content [64-66] have been shown to affect microhardness. However, our research suggests that teeth without cola immersion did not show a significant change in microhardness, in contrast with teeth treated with cola immersion, for which microhardness value decreased after the bleaching process, although there was only a slight change. This was probably because the contact period between the cola solution and the teeth over 7 days causes a decrease in pH that had the potential to cause demineralization and erosion of the tooth structure [63]. Furthermore, cola can cause an intense brown discoloration of tooth enamel [67] due to phosphoric acid and chromogen [68]. Phosphoric acid can weaken tooth enamel, causing it to erode and change color quickly due to chromagens [68]. In this study, there was no variation in the length of cola soaking on the teeth, however, cola soaking was carried out for only 7 days. Wang et al. [69] stated that during 7 days of soaking with cola, the surface of the teeth experienced discoloration and stains.

The morphology of tooth enamel abrasion before and after bleaching was observed using SEM characterization with 500x magnification. Figure 8 shows the surface morphology of tooth enamel before and after bleaching for teeth without cola immersion. There is slight damage to the tooth enamel, as indicated by the holes in the tooth enamel, as shown in Figure 8 (c). It shows that the free radicals produced by the TiO<sub>2</sub> nanocatalyst and 3% H<sub>2</sub>O<sub>2</sub> slightly damaged tooth enamel, affecting tooth hardness. The surface morphology of tooth enamel for cola soaking before bleaching in Figure 9(a) shows tooth enamel erosion in the form of holes and scratches in the tooth area. After the bleaching procedure (Figure 9(b-c)), it is clear that there are sharp scratches on the teeth and tooth enamel is damaged, which could have an impact on the tooth microhardness value.



Figure 8. Surface morphology of tooth enamel without cola soaking treatment (a) before bleaching (b) after bleaching with 0.05 g tooth whitening gel (c) after bleaching with 0.25 g tooth whitening gel



**Figure 9.** Surface morphology of tooth enamel soaked in cola (a) before bleaching (b) after bleaching with 0.05 g tooth whitening gel (c) after bleaching with 0.25 g tooth whitening gel

In ou research, different teeth were used for the treatmentwithout and with cola soaking, and the results of the Micro-Vickers test showed differences in microhardness. It should be noted that this research was focused only on the effects of bleaching using tooth whitening gel on the brightness and microhardness of teeth without and with cola soaking. Each tooth of the same type and treatment was observed for changes in color and hardness value three times after bleaching. The stability of tooth brightness after bleaching in this study has yet to be studied and the study was only focuses on the effects of bleaching on tooth brightness. In subsequent research, the stability of tooth brightness by repeatedly immersing the dye will be examined. Dehydration of teeth after bleaching can occur. The use of high  $H_2O_2$  concentrations of around 25% produces faster tooth whitening [69] but can cause high thermal sensitivity [70], resulting in more dehydration [71, 72]. Lee *et al.* [73] stated that using a low  $H_2O_2$  concentration, namely 3 to 4%, could minimize the level of thermal sensitivity or even cause no thermal sensitivity after the bleaching procedure, which could reduce dehydration. Therefore, using a low concentration as in this study, i.e. 3%  $H_2O_2$ , may not impact thermal sensitivity and may reduce tooth dehydration.

#### 4. Conclusions

PDA and chitosan-modified  $TiO_2$  samples were successfully fabricated in this study and had the following characteristics. The diffraction pattern of PDA and chitosan-modified  $TiO_2$  samples showed no change in  $TiO_2$  crystallinity, and the anatase phase was identified. Each absorption peak from the FTIR results shows the characteristics of PDA and chitosan-modified  $TiO_2$ . TEM image shows a core-shell structure where the  $TiO_2$  particles are covered with PDA and chitosan. The 3%

 $H_2O_2$  addition to the TiO<sub>2</sub> samples, modified by polydopamine and chitosan, made the material into a gel that could be used as a teeth-whitening agent. The results of three bleaches (each of 15 min) on the tooth surface with a duration under visible light irradiation over a wavelength of 420-480 nm showed that the gel was effective on brightening teeth and did not cause a significant change in the microhardness value of the teeth either with or without cola immersion. For the 0.25 g sample, the PDA-modified TiO<sub>2</sub> was found to produce tooth brightness and tooth microhardness values for teeth without cola immersion of 121.77 and 16.13 HV, respectively, while for teeth with cola immersion, the brightness and hardness values were 121.25 and 25.20 HV, respectively. In our study, a greater concentration of bleach, delivered as a greater content of PDA-modified TiO<sub>2</sub>, increased the brightness of teeth, even though  $H_2O_2$  was use at a relatively low concentration.

## 5. Acknowledgements

The researchers would like to thank the Laboratory of Materials Physics, Universitas Negeri Surabaya, for all the facilities and infrastructure for research activities ranging from TiO<sub>2</sub> synthesis and fabrication of teeth whitening gel to the preparation of microhardness tests. The researchers also thank the funders from DRTPM (Direktur Riset, Teknologi, dan Pengabdian kepada Masyarakat) based on the Decree of the Chancellor of Universitas Negeri Surabaya Number: 948/UN38/HK/KP/2023.

# References

- Sun, L., Liang, S., Sa, Y., Wang, Z., Ma, X., Jiang, T. and Wang, Y., 2011. Surface alteration of human tooth enamel subjected to acidic and neutral 30% hydrogen peroxide. *Journal of Dentistry*, 39(10), 686-692, http://doi.org/10.1016/j.jdent.2011.07.011.
- [2] Meireles, S.S., Fontes, S.T., Coimbra, L.A.A., Bona, A.D. and Demarco, F.F., 2012. Effectiveness of different carbamide peroxide concentrations used for tooth bleaching: an in vitro study. *Journal of Applied Oral Science*, 20, 186-191, https://doi.org/10.1590/S1678-77572012000200011.
- [3] De Geus, J.L., Wambier, L.M., Kossatz, S., Loguercio, A.D. and Reis, A., 2016. At-home vs in-office bleaching: a systematic review and meta-analysis. *Operative Dentistry*, 41(4), 341-356, https://doi.org/10.2341/15-287-LIT.
- [4] Mushashe, A.M., Coelho, B.S., Garcia, P.P., da Cunha, L.F., Correr, G.M. and Gonzaga, C.C., 2018. Effect of different bleaching protocols on whitening efficiency and enamel superficial microhardness. *Journal of Clinical and Experimental Dentistry*, 10(8), https://doi.org/10.4317/jced.54967.
- [5] Ghanbarzadeh, M., Ahrari, F., Akbari, M. and Hamzei, H., 2015. Microhardness of demineralized enamel following home bleaching and laser-assisted in office bleaching. *Journal of Clinical and Experimental Dentistry*, 7(3), https://doi.org/10.4317/jced.51705.
- [6] Giannini, M., Silva, A.P., Cavalli, V. and Leme, A.F.P., 2006. Effect of carbamide peroxide-based bleaching agents containing fluoride or calcium on tensile strength of human enamel. *Journal of Applied Oral Science*, 14, 82-87, https://doi.org/10.1590/S1678-77572006000200004.
- [7] Dantas, A.A.R., Bortolatto, J.F., Roncolato, A., Merchan, H., Floros, M.C., Kuga, M.C. and Junior, O.B.O., 2015. Can a bleaching toothpaste containing Blue Covarine demonstrate the same bleaching as conventional techniques? An *in vitro*, randomized and blinded study. *Journal* of Applied Oral Science, 23(6), 609-613, https://doi.org/10.1590/1678-775720150268.

- [8] Yadav, S., 2017. Bleaching effectiveness and tooth sensitivity of inoffice hydrogen peroxide containing titanium dioxide based bleaching agent: a systematic review. *Research and Reviews: Journal of Dental Sciences*, 5(1), 96-101.
- [9] Menezes, R.P.D., Silva, P.D., Leal, P.C. and Faria-e-Silva, A.L., 2018. Impact of 35% hydrogen peroxide on color and translucency changes in enamel and dentin. *Brazilian Dental Journal*, 29(1), 88-92, https://doi.org/10.1590/0103-6440201801553.
- [10] Komatsu, O., Nishida, H., Sekino, T. and Yamamoto, K., 2014. Application of titanium dioxide nanotubes to tooth whitening. *Nano Biomedicine*, 6(2), 63-72.
- [11] Istiqomah, I., Putri, A., Patmawati, T., Rohmawati, L. and Setyarsih, W., 2019. Ekstraksi titanium dioksida (TiO<sub>2</sub>) anatase menggunakan metode leaching dari pasir mineral tulungagung. *Akta Kimia Indonesia*, 4(2), 145-151. (in Indonesian)
- [12] Rohmawati, L., Istiqoah, Pratama, A.A., Setyarsih, W., Putri, N.P., Munasir and Darminto, 2022. The characteristics of TiO<sub>2</sub> anatase from Tulungagung sand as an antibacterial material. *Nanosystems: Physics, Chemistry, Mathematics*, 13(6), 640-648, https://doi.org/10.17586/2220-8054-2022-13-6-640-648.
- [13] Lalasari L.H., Firdiyono F., Yuwono A.H., Harjanto S. and Suharno, 2012. Preparation, decomposition and Ccharacterizations of Bangka-Indonesia Ilmenite (FeTiO<sub>3</sub>) derived by hydrothermal method using concentrated NaOH solution. *Advanced Materials Research*, 535-537, 750-756.
- [14] Ginting, L.I.B., Manaf, A., Astuti, W., Supriyatna, Y.I. and Bahfie, F., 2023. Study of titanium dioxide (TiO<sub>2</sub>) extraction process from Ilmenite Banten. *IOP Conference Series: Earth and Environmental Science*, 1201, https://doi.org/10.1088/1755-1315/1201/1/012092.
- [15] Ozcetin, H.K. and Surmelioglu, D., 2020. Effects of bleaching gel containing TiO<sub>2</sub> and chitosan on tooth surface roughness, microhardness and colour. *Australian Dental Journal*, 65(4), 269-277, https://doi.org/10.1111/adj.12786.
- [16] Rani, M. and Shanker, U., 2020. Green synthesis of TiO<sub>2</sub> and its photocatalytic activity. In: C.M. Hussain and A.K. Mishra, eds. *Handbook of Smart Photocatalytic Materials*. Amsterdam: Elsevier Inc, pp. 11-61.
- [17] Tomás-Gamasa, M. and Mascareñas, J.L., 2020. TiO<sub>2</sub>-based photocatalysis at theiInterface with biology and biomedicine. *ChemBioChem*, 21(3), 294-309, https://doi.org/10.1002/cbic.201900229.
- [18] Hamza, H.S.E.D., Hassan, S. and Mosallam, O., 2021. Effect of an in-office bleaching agent containing two different concentrations of titanium dioxide nano particles on the color and surface roughness of enamel. *Egyptian Dental Journal*, 67(1), 809-816, https://doi.org/ 10.21608/EDJ.2020.47230.1303.
- [19] Torres, C.R.G., Wiegand, A., Sener, B. and Attin, T., 2010. Influence of chemical activation of a 35% hydrogen peroxide bleaching gel on its penetration and efficacy—*In vitro* study. *Journal of Dentistry*, 38(10), 838-846, https://doi.org/10.1016/j.jdent.2010.07.002.
- [20] Kishi, A., Otsuki, M., Sadr, A., Ikeda, M. and Tagami, J., 2011. Effect of light units on tooth Bleaching with visible-light activating titanium dioxide photocatalyst. *Dental Materials Journal*, 30(5), 723-729, https://doi.org/10.4012/dmj.2010-210.
- [21] Brenneisen, P., Sies, H. and Scharffetter-Kochanek, K., 2002. Ultraviolet-B irradiation and matrix metalloproteinases. *Annals of the New York Academy of Sciences*, 973(1), 31-43, https://doi.org/10.1111/j.1749-6632.2002.tb04602.x.
- [22] Ebersole, J.L., Kirakodu, S., Novak, M.J., Stromberg, A.J., Shen, S., Orraca, L., Gonzalez-Martinez, J., Burgos, A. and Gonzales, O.A., 2014. Cytokine gene expression profiles during initiation, progession and resolution of periodontitis. *Journal of Clinical Periodontology*, 41, 853-861, https://doi.org/10.1111/jcpe.12286.
- [23] Luong, M.N., Otsuki, M., Shimada, Y., Ei, T.Z., Sumi, Y. and Tagami, J., 2018. Effect of lights with various wavelengths on bleaching by 30% hydrogen peroxide. *Laser in Medical Science*, 34, 901-906, https://doi.org/10.1007/s10103-018-2670-y.

- [24] Zhang, F., Wu, C., Zhou, Z., Wang, J., Bao, W., Dong, L., Zhang, Z., Ye, J., Liao, L. and Wang, X., 2018. Blue-light-activated nano-TiO<sub>2</sub>@PDA for highly effective and nondestructive tooth whitening. ACS Biomaterials Science and Engineering, 4(8), 3072-3077, https://doi.org/10.1021/acsbiomaterials.8b00548.
- [25] Suemori, T., Kato, J., Nakazawa, T., Akashi, G. and Hirai, Y., 2008. A new non-vital tooth bleaching method using titanium dioxide and 3.5% hydrogen peroxide with a 405-nm diode laser or a halogen lamp. *Laser Physics Letters*, 5(6), 454-459.
- [26] Sun, X., Yan, L., Xu, R., Xu, M. and Zhu, Y., 2019. Surface modification of TiO<sub>2</sub> with polydopamine and its effect on photocatalytic degradation mechanism. *Colloids and Surfaces A: Physicochemical and Engineering Aspects*, 570, 199-209.
- [27] Khan, Z., Shanker, R., Um, D., Jaiswal, A. and Ko, H., 2018. Bioinspired polydopamine and composites for biomedical applications. In: A. Khan, M. Jawaid, A.A.P. Khan and A.M. Asiri, eds. *Electrically Conductive Polymer and Polymer Composites: From Synthesis to Biomedical Applications*. New York: John Wiley and Sons Inc, pp. 1-29.
- [28] Liebscher, J., Mrówczyński, R., Scheidt, H.A., Filip, C., Hădade, N.D., Turcu, R., Bende, A. and Beck, S., 2013. Structure of polydopamine: a never-ending story? *Langmuir*, 29(33), 10539-10548, https://doi.org/10.1021/la4020288.
- [29] Muşat, V., Anghel, E.M., Zaharia, A., Atkinson, I., Mocioiu, O.C., Buşilă, M. and Alexandru, P., 2021. A chitosan–agarose polysaccharide-based hydrogel for biomimetic remineralization of dental enamel. *Biomolecules*, 11(8), https://doi.org/10.3390/biom11081137.
- [30] Pini, N.I.P., Piccelli, M.R., Vieira-Junior, W.F., Ferraz, L.N., Aguiar, F.H.B. and Lima, D.A.N.L., 2022. In-office tooth bleaching with chitosan-enriched hydrogen peroxide gels: *In vitro* results. *Clinical Oral Investigations*, 26(1), 471-479, https://doi.org/10.1007/s00784-021-04021-4.
- [31] Hamman, J.H., 2010. Chitosan based polyelectrolyte complexes as potential carrier materials in drug delivery systems. *Marine Drugs*, 8(4), 1305-1322.
- [32] Feng, Y. and Xia, W., 2011. Preparation, characterization and antibacterial activity of watersoluble O-fumaryl-chitosan. *Carbohydrate Polymers*, 83(3), 1169-1173.
- [33] Arnaud, T.M.S., de Barros Neto, B. and Diniz, F.B., 2010. Chitosan effect on dental enamel de-remineralization: an *in vitro* evaluation. *Journal of Dentistry*, 38(11), 848-852, https://doi.org/10.1016/j.jdent.2010.06.004.
- [34] Podust, T.V., Kulik, T.V., Palyanytsya, B.B., Gun'Ko, V.M., Tóth, A., Mikhalovska, L., Menyhárd, A. and László, K., 2014. Chitosan-nanosilica hybrid materials: preparation and properties. *Applied Surface Science*, 320, 563-569, https://doi.org/10.1016/j.apsusc.2014.09.038.
- [35] Li, H., Du, Y., Xu, Y., Zhan, H. and Kennedy, J.F., 2004. Interactions of cationized chitosan with components in a chemical pulp suspension. *Carbohydrate Polymers*, 58(2), 205-214, https://doi.org/10.1016/j.carbpol.2004.06.044.
- [36] Pratama, A.A. and Rohmawati, L., 2021. Analisis kristalinitas TiO<sub>2</sub>@PDA hasil leaching ilmenite tulungagung. *Inovasi Fisika Indonesia*, 10(2), 68-72, https://doi.org/10.26740/ifi.v10n2.p68-72. (in Indonesian)
- [37] Almejrad, L., Almansour, A., Bartlett, D. and Austin, R., 2024. CAD/CAM leucite-reinforced glass-ceramic for simulation of attrition in human enamel in vitro. *Dental Materials*, 40, 173-178, https://doi.org/10.1016/j.dental.2023.11.004.
- [38] Saranya, K.S., Padil, V.V.T., Senan, C., Pilankatta, R., Saranya, K., George, B., Wacławek, S. and Černík, M., 2018. Green synthesis of high temperature stable anatase titanium dioxide nanoparticles using gum kondagogu: characterization and solar driven photocatalytic degradation of organic dye. *Nanomaterials*, 8(12), https://doi.org/10.3390/nano8121002.
- [39] Kalaiarasi, S. and Jose, M., 2017. Dielectric functionalities of anatase phase titanium dioxide nanocrystals synthesized using water-soluble complexes. *Applied Physics A*, 123(8), https://doi.org/10.1007/s00339-017-1121-0.

- [40] Habiba, U., Joo, T.C., Shezan, S.K.A., Das, R., Ang, B.C. and Afifi, A.M., 2019. Synthesis and characterization of chitosan/TiO<sub>2</sub> nanocomposite for the adsorption of Congo red. *Desalination and Water Treatment*, 164, 361-367, https://doi.org/10.5004/dwt.2019.24391.
- [41] Abisharani, J.M., Devikala, S., Kumar, R.D., Arthanareeswari, M. and Kamaraj, P., 2019. Green synthesis of TiO<sub>2</sub> nanoparticles using *Cucurbita pepo* seeds extract. *Materials Today: Proceedings*, 14(Part 2), 302-307, https://doi.org/10.1016/j.matpr.2019.04.151.
- [42] Al-Zahrani, H., El-Waseif, A. and El-Ghwas, D.E., 2018. Biosynthesis and evaluation of TiO<sub>2</sub> and ZnO nanoparticles from in vitro stimulation of *Lactobacillus johnsonii*. Journal of Innovations in Pharmaceutical and Biological Sciences, 5(1), 16-20.
- [43] Żeglin, J., Piotrowski, G.P. and Piękos, R., 2006. A study of interaction between hydrogen peroxide and silica gel by FTIR spectroscopy and quantum chemistry. *Journal of Molecular Structure*, 794(1-3), 83-91, https://doi.org/10.1016/j.molstruc.2006.01.043.
- [44] Queiroz, M.F., Teodosio Melo, K.R., Sabry, D.A., Sassaki, G.L. and Rocha, H.A.O., 2014. Does the use of chitosan contribute to oxalate kidney stone formation? *Marine Drugs*, 13(1), 141-158, https://doi.org/10.3390/md13010141.
- [45] Singh, L.K. and Koiry, B.P., 2018. Natural dyes and their effect on efficiency of TiO<sub>2</sub> based DSSCs: a comparative study. *Materials Today: Proceedings*, 5(1), 2112-2122, https://doi.org/10.1016/j.matpr.2017.09.208.
- [46] Saravanan, S. and Dubey, R.S., 2021. Optical and morphological studies of TiO<sub>2</sub> nanoparticles prepared by sol-gel method. *Materials Today: Proceedings*, 47(9), 1811-1814, https://doi.org/10.1016/j.matpr.2021.03.207.
- [47] Cui, J., Ma, C., Li, Z., Wu, L., Wei, W., Chen, M., Peng, B. and Deng, Z., 2016. Polydopaminefunctionalized polymer particles as templates for mineralization of hydroxyapatite: biomimetic and in vitro bioactivity. *RSC Advances*, 6(8), 6747-6755, https://doi.org/10.1039/C5RA24821C.
- [48] Rahoui, N., Hegazy, M., Jiang, B., Taloub, N. and Huang, Y.D., 2018. Particles size estimation of polydopamine based polymeric nanoparticles using near-infrared spectroscopy combined with linear regression method. *American Journal of Analytical Chemistry*, 9(5), 273-285, https://doi.org/10.4236/ajac.2018.95021.
- [49] Jeon, W.-Y., Kim, H.-H. and Choi, Y.-B., 2021. Development of a glucose sensor based on glucose dehydrogenase using polydopamine-functionalized nanotubes. *Membranes*, 11(6), https://doi.org/10.3390/membranes11060384.
- [50] Ghorbani, F., Zamanian, A. and Torabinejad, B., 2020. The effect of oxygen plasma pretreatment on the properties of mussel-inspired polydopamine-decorated polyurethane nanofibers. *Journal* of *Polymer Engineering*, 40(2), 109-119, https://doi.org/10.1515/polyeng-2019-0219.
- [51] Chen, F., Yu, W., Qie, Y., Zhao, L., Zhang, H. and Guo, L.H., 2019. Enhanced photocatalytic removal of hexavalent chromium through localized electrons in polydopamine-modified TiO<sub>2</sub> under visible irradiation. *Chemical Engineering Journal*, 373, 58-67, https://doi.org/10.1016/j.cej.2019.05.022.
- [52] Ahghari, M.A., Ahghari, M.R., Kamalzare, M. and Maleki, A., 2022. Design, synthesis, and characterization of novel eco-friendly chitosan-AgIO<sub>3</sub> bionanocomposite and study its antibacterial activity. *Scientific Reports*, 12, https://doi.org/10.1038/s41598-022-14501-6.
- [53] Hussein, E.M., Desoky, W.M., Hanafy, M.F. and Guirguis, O.W., 2021. Effect of TiO<sub>2</sub> nanoparticles on the structural configurations and thermal, mechanical, and optical properties of chitosan/TiO<sub>2</sub> nanoparticle composites. *Journal of Physics and Chemistry of Solids*, 152, https://doi.org/10.1016/j.jpcs.2021.109983.
- [54] Tanno, Y., Otsuki, M., Nishimura, M., Luong, M.N., Takagaki, T., Nakajima, M., Sumi, Y. and Tagami, J., 2020. Effect of ultraviolet ray on tooth bleaching using titanium dioxide photocatalyst. *Asian Pacific Journal of Dentistry*, 20(2), 35-40, https://doi.org/10.47416/apjod.20-0277.
- [55] Razali, M.H., Dris, M.R.M. and Rudin, N.N.S.M., 2009. Photodegradation of methyl orange dye using titanium dioxide photocatalyst. *Journal of Sustainability Science and Management*, 4(1), 49-55.

- [56] Garcia, J.C. and Takashima, K.J, 2003. Photocatalytic degradation of imazaquin in an aqueous suspension of titanium dioxide. *Journal of Photochemistry and Photobiology A: Chemistry*, 155(1-3), 215-222, https://doi.org/10.1016/S1010-6030(02)00370-2.
- [57] Chen, S. and Liu, Y., 2007. Study on the photocatalytic degradation of glyphosate by TiO<sub>2</sub> photocatalyst. *Chemosphere*, 67(5), 1010-1017, https://doi.org/10.1016/j.chemosphere.2006.10.054.
- [58] Kodir, A.I.A., 2014. Teknik bedah dengan skalpel pada hiperpigmentasi gingiva. Odonto Dental Journal, 1(2), 40-45. (in Indonesian)
- [59] Jiang, Y., Shi, K., Tang, H. and Wang, Y., 2019. Enhanced wettability and wear resistance on TiO2/PDA thin films prepared by sol-gel dip coating. *Surface and Coatings Technology*, 375, 334-340, https://doi.org/10.1016/j.surfcoat.2019.07.051.
- [60] Zanin, F., 2016. Recent advances in dental bleaching with laser and LEDs. *Photomedicine and Laser Surgery*, 34(4), 135-136, https://doi.org/10.1089/pho.2016.4111.
- [61] Lugo-Varillas, J.G., Tinedo-López, P.L., Watanabe, O.G., Correa, M.A., Álvarez, V.E. and Hermoza, N.M., 2020. Influence of pH value of bleaching gels on surface roughness of bovine enamel. *Odovtos-International Journal of Dental Sciences*, 22(2), 113-123.
- [62] Klaric, E., Rakic, M., Sever, I., Milat, O., Par, M. and Tarle, Z., 2015. Enamel and dentin microhardness and chemical composition after experimental light-activated bleaching. *Operative Dentistry*, 40(4), E132-E141, https://doi.org/10.2341/14-148-L.
- [63] Bistey, T., Nagy, I.P., Simó, A. and Hegedűs, C., 2007. In vitro FT-IR study of the effects of hydrogen peroxide on superficial tooth enamel. Journal of Dentistry, 35(4), 325-330, https://doi.org/10.1016/j.jdent.2006.10.004.
- [64] Rodrigues, J.A., Oliveira, G.P.F. and Amaral, C.M., 2007. Effect of thickener agents on dental enamel microhardness submitted to at-home bleaching. *Brazilian Oral Research*, 21(2), 170-175, https://doi.org/10.1590/S1806-83242007000200013.
- [65] Kwon, S.R., Kurti, S.R., Oyoyo, U. and Li, Y., 2015. Effect of various tooth whitening modalities on microhardness, surface roughness and surface morphology of the enamel. *Odontology*, 103, 274-279, https://doi.org/10.1007/s10266-014-0163-4.
- [66] Sharma, A., Anggarwal, N., Rastogi, S., Choudhury, R. and Tripathi, S., 2017. Effectiveness of platelet-rich fibrin in the management of pain and delayed wound healing associated with established alveolar osteitis (dry socket). *European Journal of Dentistry*, 11(4), 508-513, https://doi.org/10.4103/ejd.ejd\_346\_16.
- [67] Hanin, S. and Thenmozhi, M.S., 2018. Association between dental erosion and carbonated drinks. *Drug Invention Today*, 10(11), 2335-2337.
- [68] Sarembe, S., Kiesow, A., Pratten, J., Webster, C., 2022. The impact on dental staining caused by beverages in combination with chlorhexidine digluconate. *European Journal of Dentistry*, 16(4), 911-918, https://doi.org/10.1055/s-0041-1742123.
- [69] Wang, Y., Wen, X., Jia, Y., Huang, M., Wang, F., Zhang, X., Bai, Y., Yuan, G. and Wang, Y., 2020. Piezo-catalysis for nondestructive tooth whitening. *Nature Communications*, 11, 1-11, https://doi.org/10.1038/s41467-020-15015-3.
- [70] Papathanasiou, A., Kastali, S., Perry, R.D. and Kugel, G., 2002. Clinical evaluation of a 35% hydrogen peroxide in-office whitening system. *Compendium of Continuing Education in Dentistry*, 23, 335-338.
- [71] Yarborough, D.K., 1991. The safety and efficacy of tooth bleaching: a review of the literature 1988-1990. *Compendium*, 12(3), 191-196.
- [72] Kuliński, W. and Mróz, J., 2023. Physical management in Algodystrophic syndrome. A clinical study. Acta Balneologica, 5(177), 271-276, https://doi.org/10.36740/ABAL202305101.
- [73] Lee, Y., Kwon, S. -K. and Park, J.-W., 2008. The effectiveness of sealing technique on in-office bleaching. *The Journal of Korean Academy of Conservative Dentistry*, 33(5), 463-471, https://doi.org/10.5395/JKACD.2008.33.5.463.