# สัตวแพทย์มหานครสาร

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# ไวรัสฝีดาษกับการเกิดโรคไฟโบรมาโทซีสที่ผิวหนังในกระบือเผือก: รายงานสัตว์ป่วย

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**บทคัดย่อ:** ตัวอย่างชิ้นเนื้อผิวหนังจากกระบือเผือกเพศผู้อายุ 5 ปี ได้ถูกส่งเข้ารับการตรวจทางจุลพยาธิวิทยา จากประวัติการพบก้อนที่ผิวหนังหลายบริเวณร่วมกับเป็นแผลหลุมและสะเก็ดมานาน 1 ถึง 2 เดือน ผลการ ตรวจทางจุลพยาธิวิทยา พบการเกิดแผลหลุมร่วมกับการอักเสบบริเวณด้านบนของผิวหนัง ด้านล่างพบการ เพิ่มจำนวนของเซลล์ไฟโบรบลาสท์โดยแทรกอยู่ระหว่างมัดของคอลลาเจน พบอินคลูชันบอดีในไซโทพลาซึม ของไฟโบรบลาสท์ลักษณะกลมรีติดสีชมพู การตรวจทางอิมมูโนฮีสโตเคมีให้ผลบวกอย่างเด่นชัดต่อโปรตีนโครง ร่างชนิดไวเมนติน แต่ให้ผลลบทางอิมมูโนฮีสโตเคมีต่อโปรตีนโครงร่างชนิดแพนไซโทเคอราทีน จากข้อมูล ข้างต้นสรุปได้ว่าเกิดโรคไฟโบรมาโทซีสที่ผิวหนัง การตรวจทางกล้องจุลทรรศน์อิเลคตรอนชนิดลำแสงส่องผ่าน โดยใช้เนื้อเยื่อที่ดองไว้ในฟอร์มาลิน พบเชื้อไวรัสที่ยังเจริญไม่เต็มที่ขนาดประมาณ 200 x 300 นาโนเมตร ลักษณะคล้ายพ๊อกไวรัสภายในบริเวณที่เป็นก้อนอินคลูชั้นบอดี โรคไฟโบรมาโทซีสที่มีการตรวจพบอินคลูชั้น บอดี มีรายงานพบน้อยในคนและสัตว์ สำหรับในสัตว์ พ็อกไวรัสมีรายงานก่อโรคไฟโบรมาโทซีสและมึกโซมาโท ซีสในกระต่ายบ้านและกระต่ายป่า จากการสืบค้นข้อมูล ไม่การรายงานว่าพ๊อกไวรัสเกี่ยวข้องกับการเกิดไฟโบร มาโทซีสชนิดที่พบอินคลูชันบอดีในไซโทพลาซึมในกระบือเผือกมาก่อน รายงานฉบับนี้ เป็นการรายงานรูปแบบ ที่ไม่พบบ่อยของการติดเชื้อพ๊อกไวรัส โดยที่พบลักษณะรอยโรคเป็นแบบไฟโบรมาโทซีสในกระบือเผือก สัตว แพทย์ที่ตรวจวินิจฉัยกระบือที่พบรอยโรคเป็นก้อนที่ผิวหนังและมีแผลหลุมหลายบริเวณโดยไม่ประสบผลสำเร็จ ในการรักษา ควรคำนึงถึงโอกาสเกิดโรคไฟโบรมาโทซีสจากการติดเชื้อพ็อกไวรัส เช่นเดียวกับในกรณีศึกษานี้ ด้วย

คำสำคัญ: กระบือ ผิวหนัง เผือก พยาธิวิทยา ไฟโบรมาโตซีส ไวรัสฝีดาษ

#ผู้รับผิดชอบบทความ

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Poxvirus Associated with Cutaneous Fibromatosis in an Albino Buffalo (*Bubalus bubalis*): A Case Report

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Abstract: Skin biopsy of a five-year-old male albino buffalo (Bubalus bubalis) was submitted for histopathological diagnosis. The buffalo had a history of multiple skin nodules, ulceration and scab formation at the trunk for about two months. Histopathological examination revealed superficial ulceration with inflammatory reaction and underlying proliferation of fibroblasts among collagen bundles within the dermis. The proliferated fibroblasts showed large oval to round eosinophilic intracytoplasmic inclusion bodies. Immunohistochemically (IHC), the proliferated cells were strongly positive to vimentin. Immunohistochemical reaction to pancytokeratin was negative. The diagnosis of cutaneous fibromatosis was made. For transmission electron microscopic examination from the formalin fixed skin samples, some immature virions, about 200x300 nm. in size and morphologic consistent with poxvirus, were detected within cytoplasmic inclusion body. Fibromatosis with intracytoplasmic inclusion body was a rare condition reported in human and animals. In animals, poxvirus associated with fibromatosis or myxomatosis was well documented in Lagomorphs such as rabbits and hares. The presence of poxvirus associated with fibromatosis with eosinophilic intracytoplasmic inclusion body in an albino buffalo has never been documented in the literature. This article demonstrates atypical form of poxvirus infection with histopathologically appearance as fibromatosis in an albino buffalo. Veterinarians deal with nodular and ulcerative skin lesions without responsive to treatment in buffalo should keep in mind of a possible fibromatosis, poxviral associated lesions, as observed in this study.

Keywords: Albino, Buffalo, Fibromatosis, Pathology, Poxvirus, Skin

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# Introduction

Cutaneous fibromatosis is a condition with multiple nodules of fibroblastic proliferation has been found on skin. However, fibromatosis with intracytoplasmic inclusion body is a rare condition reported in human and animals. Poxvirus is one of the common causes of fibroblastic most proliferation in animals. The shape of Poxviral virions is oval or brick-shape of about 200 to 400 nm in length, with axial ratios of 1.2 to forms 1.7. There are two of the nonenveloped virion namely the intact M (mulberry) form and the C (capsule) form (Mark and Palumbo, 1991). A well-known Leporipoxvirus induced fibroma and fibromatosis is the rabbit (shope) fibroma virus and squirrel fibroma virus (Bangari et al., 2009). This virus causes fibroma on the legs, feet, and ears and persists for several months. Intracytoplasmic inclusion bodies can be histologically detected in fibroblasts. Other example of virus induced fibromatosis is papillomavirus infection in roe deer and other animals of family Cervidae such as whitetailed deer, red deer, mule deer, European elk and reindeer (Kureljušić et al., 2012; Sunderberg and Nielsen, 1981). Equine sarcoid is an example of locally aggressive, fibroblastic skin tumor of animals in family Equidae such as horses, mules, donkeys and zebra (Jangir et al., 2013). Bovine

papillomavirus (BPV-1 and BPV-2) has been reported to be associated with equine sarcoid pathogenesis (Chambers *et al.*, 2003; Bogaert *et al.*, 2008; Kumar *et al.*, 2013). Although papillomaviruses have been described as highly species—specific, however, interspecies infection occurs as in equine sarcoid (Jangir *et al.*, 2013). Buffalopox virus infection is a proliferative viral skin disease occurs in buffalos, cows and human. In a recent outbreak of this virus in India did not demonstrate any fibroblastic proliferation (Venkatesan *et al.*, 2009).

The aim of this study is to describe histopathological, immunohistochemical and transmission electron microscopical findings of a rare condition of cutaneous fibromatosis in an albino buffalo (*Bubalus bubalis*).

# Materials and methods

Case history: A five-year-old male albino buffalo (Bubalus bubalis) with a history of multiple skin nodules, ulceration and scab formation at trunk for about two months without satisfactory treatment result. Skin biopsy was performed and the specimen was submitted in 10 % phosphate-buffered formalin to Mahanakorn Veterinary Diagnostic Center for histopathological investigation.

*Histopathology*: Tissue slides for microscopic examination were prepared using routine

paraffin embedding technique. Briefly, the tissues were processed and embedded in paraffin, sectioned at 4 microns and stained with hematoxylin and eosin (H&E).

*Immunohistochemistry* (IHC): For immunohistochemistry, the polymer detection system (NovoLink Cat. No. RE7140-K, Chemo Science) was applied. **Primary** antibodies to pancytokeratin (dilution 1:400; AE1/AE3, Nichirei) and vimentin (dilution 1:100: Clone V9, Nichirei) were used with 3-3'diaminobenzidine (DAB, Dako) as chromogen as previously described (Ramos-Vara et al., 2003; Mamom, 2012). Briefly, the tissue sections were deparaffinized and hydrated. Antigen retrieval was performed by boiling tissue sections in citrate buffer using Inhibition endogenous microwave. of peroxidase was done using hydrogenperoxide. Other steps including the use of blocking protein, primary antibodies, secondary antibody, peroxidase enzyme, substrate/chromogen and hematoxylin counterstain were done according to the product information (NovoLink Cat. No. RE7140-K, Chemo Science). The slides were evaluated under light microscope.

Transmission Electronmicroscopy (TEM):

Specimen preparation for transmission electron microscopic examination using

formalin fixed tissue was done at Pathology Section, the National Institute of Animal Health (NIAH) according to the previous study (Mamom *et al.*, 2010). Briefly, the tissue samples were embedded in epoxy resin (Epon). The ultrathin sections were cut and stained with 5% uranyl acetate and lead citrate. The samples were examined under transmission electron microscope (JEM-1200EX, JEOL, Japan).

# Results

# Histopathological findings:

Histopathologic examination of hematoxylin & eosin (H&E) stained slides revealed fibroblastic proliferation among preexisting dermal collagen fibers (Fig. 1A) with expansive growth to superficial dermis and extended deep into subcutis. proliferated fibroblasts showed oval nuclei with less heterochromatin and prominent nucleoli. Large oval to round shaped eosinophilic intracytoplasmic inclusion bodies (Fig. 1B) were found in many cells fibroblasts. Some underwent cytoplasmic vacuolar change. Mitotic figures were rare and ranged from 0 to 2 per high of field. Areas necrosis power hemorrhages were seen especially at the center of nodules. The superficial epidermis revealed necrosis, ulceration and hemorrhage with inflammatory cells infiltration especially neutrophils. No epithelial cell proliferation and ballooning degeneration were observed from the submitted samples.

# Immunohistochemical findings:

Vimentin and pancytokeratin are cytoskeletal proteins normally observed in cytoplasm of the cells of mesenchymal and epithelial origin respectively. The proliferated fibroblasts in this case expressed vimentin (Fig. 1C). Immunoreactivity to pancytokeratin was negative. This result confirmed the non-epithelial origin of proliferative cells. The intracytoplasmic inclusion bodies were also strongly positive for vimentin as well.

# Transmission electron microscopic (TEM) findings:

Transmission electron microscopic (TEM) findings results showed some immature virions of about 200 to 300 nm. in size (Fig. 1D) in the area of cytoplasmic inclusion bodies. The virions contained dumbell-shaped nucleocapsid characteristic for poxvirus.

### Discussion

The diagnosis of fibromatosis in this case was done on the basis of gross and microscopic lesions and immuno histochemical results. The presence of poxvirus within the cytoplasmic inclusion bodies by transmission electron microscopic

confirmed the examination cause fibroblastic proliferation in this case (Mark and Palumbo, 1991; Fenner and Nakano, 1988; Westwood et al., 1964). The absence of epithelial lesions, especially ballooning degeneration, is possibly due to the small biopsy sample which was collected from ulcerated lesion. The presence of immature virions in intracytoplasmic inclusion body verified the role of poxvirus in inducing fibroblastic proliferation in buffalo which is similar to a condition in rabbit caused by poxvirus infection, rabbit (shope) fibroma virus. Whether the albino status of this buffalo plays an important role as predisposing factor on the tumor development is unclear because of the lack of information concerning the tumorigenesis in albino animal caused by poxvirus infection. Unfortunately, the animal die and the owner did not allowed to use the carcass for further studies. The type of poxvirus in this case could not be able to identify and requires additional molecular study.

In conclusions, we reported the presence of poxvirus in association with fibromatosis in an albino buffalo (*Bubalus bubalis*), which has never been documented in the literatures. This article demonstrated atypical form of poxvirus infection with histopathological appearance as fibromatosis. Veterinarians who deal with nodular and

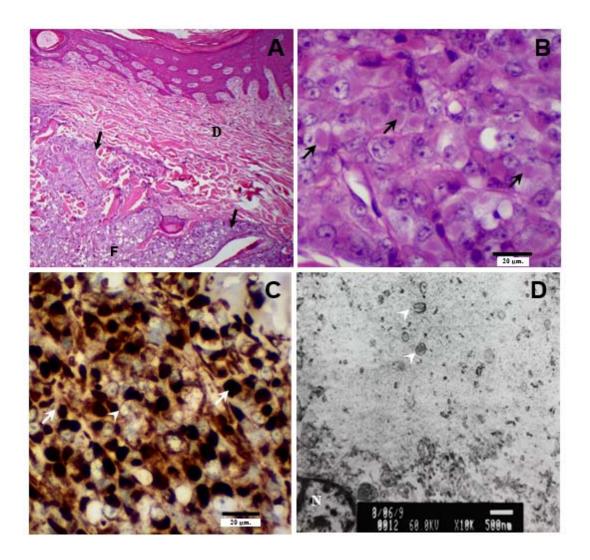


Fig. 1 A-D: Histopathological, immunohistochemical and transmission electron microscopic findings of skin lesion. 1A: Outline (black arrows) of fibroblastic proliferation (F) within dermis (D). 1B: Higher magnification of fibroblastic proliferation. Large round eosinophilic intracytoplasmic inclusion bodies (black arrows) were detected in some proliferated fibroblasts. 1C: The proliferated fibroblasts showed positive IHC to vimentin shown as brown staining area in cytoplasm. Very strong immunoreactivity was found at cytoplasmic inclusion bodies (white arrows); arrow head = nuclei of fibroblast; DAB with hematoxylin counter stain. 1D: Transmission electron microscopic picture of immature virions (white arrow heads), about 200x300 nm. in size, morphologic consistent with poxvirus. N = nucleus. Bar = 500 nm.

ulcerative skin lesions without response to treatment in buffalo should keep in mind of a possible fibromatosis, poxviral associated lesions, as observed in this study.

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#### References

- Bangari, D.S., Miller, M.A., Stevenson, G.W., Thacker, H.L., Sharma, A., and Mittal, S.K. 2009. Cutaneous and Systemic Poxviral Disease in Red (Tamiasciurus hudsonicus) and Gray (Sciurus carolinensis) Squirrels. *Vet. Pathol.* 46:667–672.
- Bogaert L., Martens A., Depoorter P. and Gasthuys F. 2008. Equine sarcoids association with bovine papillomavirus. Vlaams. Diergen. Tijdsch. 78: 131–137.
- Chambers G., Ellsmore V.A., O'Brien P.M., Reid S.W.J., Love S., Campo M.S. and Nasir, L. 2003. Association of bovine papillomavirus with the equine sarcoid. *J. Gen. Virol.* 8:1055–1062.
- Fenner, F., and Nakano, J.H. 1988. Poxviridae, p. 177-210. In E. H. Lennette, P. Halonen, and F. A. Murphy (ed.). Laboratory diagnosis of infectious diseases: principles and practice. vol. 2. Springer-Verlag, New York.
- Jangir B.L., Singh V.K., Saikumar G., Pawde A.M.,
  Churamani C.P. and Somvanshi R. 2013.
  Preliminary pathological studies on
  equine skin lesions with specific
  reference to Equine Sarcoid and

- Detection of Bovine papillomaviruses 1 and 2. *Adv. Anim. Vet. Sci.* 1 (6): 197-201.
- Kumar P., Nagarajan N., Kumar D., Bind R.B. and Somvanshi R. 2013. Detection and quantification of bovine papillomaviruses (BPVs) in cutaneous warts of cattle and buffaloes. *Adv. Amin. Vet. Sci.* 1 (2): 53–58.
- Kureljušić, B. 1, Savić, B.1, Pavlović, I. 1, Jezdimirović, N.1, Kureljušić, J.1 , Stanojević, S.1, Minić, S.2, Ivetić, V. 2012. Fibromatosis of the roe deer (*Capreolus capreolus*). International symposium on hunting, **M**odern aspects of sustainable management of game population, Zemun-Belgrade, Serbia, 22. 24. June 2012.
- Mamom, T. 2012. Metastatic Urothelial Carcinoma with Systemic Amyloidosis in a Male Pseudohermaphrodite Goat. *J. Mahanakorn Vet. Med.* 7(2): 109-118.
- Mamom, T., Dumrongsoonthornchai, P. and Trongwongsa, L. 2010. Avian Polyomavirus Infection in Non-Budgerigar Psittacine Birds in Thailand: Case Report. *Thai J. Vet. Med.* 40(1): 75-80
- Mark, R., Buller, L. and Palumbo J. Gregory, 1991. Poxvirus Pathogenesis, In Microbiological Review, Mar., p. 80-122.
- Ramos-Vara, J.A., Miller, M.A., Boucher, M., Roudabush, A. and Johnson, G.C. 2003.

  Immunohistochemical detection of

uroplakin III, cytokeratin 7, and cytokeratin 20 in canine urothelial tumors. *Vet Pathol*. 40 (1): 55-62.

Sundberg, J.P. and S.W. Nielsen. 1981. Deer Fibroma: A Review. *Can. vet. J.* 22: 385-388.

Venkatesan G., Balamuruga V., Prabhu M., Yogisharadhya R., Bora P.D., Gandhale P.N., Siva Sanka M.S., Kulkarni A.M., Singh R.K. and Bhanuprakash V. 2009. Emerging and re-emerging zoonotic buffalopox infection: a severe out break on Kolhapur (Maharashtra), India. *Veter. Ital.* 46 (4): 439-448.

Westwood, J.C.N., Harris, W.J., Zwartouw, H.T., Titmuss, D.H.J., and Appleyard, G.. 1964. Studies on the structure of vaccinia virus. *J. Gen. Virol.* 34:67-78.

