

# Perilla Mint (*Perilla frutescens*): An Alternative Animal Feed to Enhance Omega-3 Fatty Acids in Meat and Eggs as Functional Food

## งาชี่ม่อน: ทางเลือกอาหารสัตว์เพื่อเพิ่มปริมาณกรดไขมันโอเมก้าสามในเนื้อสัตว์และไข่สำหรับเป็นอาหารเพื่อสุขภาพ

Napatsorn Montha<sup>1</sup>, Winai Yothinsirikul<sup>2</sup>, Kittiphong Tippaya<sup>2</sup>, Kanchit Chompupan<sup>2</sup>,

Christian Lambertz<sup>3</sup> and Sanchai Jaturasitha<sup>1\*</sup>

ณภัสสร มณฑา<sup>1</sup> วินัย โยธินศิริกุล<sup>2</sup> กิตติพงษ์ ทิพย์ะ<sup>2</sup> ครรชิต ชมพูนันท์<sup>2</sup>

Christian Lambertz<sup>3</sup> และ สัญชัย จตุรสิทธา<sup>1\*</sup>

<sup>1</sup>Department of Animal and Aquatic Sciences, Faculty of Agriculture, Chiang Mai University, Chiang Mai 50200, Thailand

<sup>1</sup>ภาควิชาสัตวศาสตร์และสัตว์น้ำ คณะเกษตรศาสตร์ มหาวิทยาลัยเชียงใหม่ ต.สุเทพ อ.เมือง จ.เชียงใหม่ 50200

<sup>2</sup>Faculty of Animal Science and Technology, Maejo University, Chiang Mai 50290, Thailand

<sup>2</sup>ภาควิชาสัตวศาสตร์และเทคโนโลยี มหาวิทยาลัยแม่โจ้ ต.หนองหาร อ.สันทราย จ.เชียงใหม่ 50290

<sup>3</sup>Faculty of Science and Technology, Free University of Bozen-Bolzano, Bolzano, Italy

\*Corresponding author: Email: ja.sanchai@gmail.com

(Received: 6 March 2017; Accepted: 21 July 2017)

**บทคัดย่อ:** สุขภาพมนุษย์กลายเป็นเรื่องที่น่าวิตกกังวลจากตัวเลขของจำนวนผู้ป่วยที่เป็นโรคหลอดเลือดหัวใจทั่วโลกที่กำลังเพิ่มสูงขึ้น การบริโภคอาหารในลักษณะของอาหารเพื่อสุขภาพ (functional food) โดยเฉพาะอย่างยิ่ง omega-3 ซึ่งเป็นทางเลือกหนึ่งในการรักษาสุขภาพโดยการบริโภคสารอาหารที่ดีต่อสุขภาพ จากการรายงานข้อดีของกรดไขมันโอเมก้า (omega) 3, 6 และ 9 ในด้านการลดความเสี่ยงของการเกิดโรคหลอดเลือดหัวใจได้เป็นอย่างดี omega-3 เป็นกรดไขมันที่ทราบกันดีว่ามักพบในปลาทะเล แต่เมื่อไม่นานมานี้มีนักวิจัยรายงานว่าพบในงาชี่ม่อน (*Perilla frutescens*) เป็นแหล่งกรดไขมัน omega-3 ที่มีศักยภาพสูงเช่นกัน โดยเฉพาะเมล็ดงาชี่ม่อนมีการรายงานว่าพบองค์ประกอบของ omega-3 สูงและเมื่อใช้เมล็ดงาชี่ม่อนและกากงาชี่ม่อน รวมในอาหารสัตว์ เช่น สุนัข และไก่ พบว่าสามารถเพิ่ม omega-3 ในเนื้อสุกร ไก่เนื้อ และไข่ได้ และข้อดีของงาชี่ม่อนยังสามารถลดต้นทุนการผลิตสัตว์ ได้เนื้อที่มีคุณค่าสูงกว่าแหล่งโปรตีนในอาหารสัตว์ทดแทนแหล่งโปรตีนอื่นเช่น กากถั่วเหลือง นอกจากนี้เมล็ดงาชี่ม่อนยังมีสารยับยั้งการทำงานของเอนไซม์ทริปซินต่ำกว่ากากถั่วเหลืองอีกด้วย ซึ่งถือว่าเป็นข้อดีในการนำมาใช้เป็นอาหารสัตว์

**คำสำคัญ:** งาชี่ม่อน ลักษณะซาก คุณภาพเนื้อ

**Abstract:** Human health is an increasing concern reflected, for example, by a rising number of patients with diseases like coronary artery disease (CAD) worldwide. The consumption of functional food, which is rich in omega-3 fatty acid is one possibility to improve health through a better nutrition. In the literature, benefits of omega-3, -6 and -9 fatty acids in terms of reducing the risk for CAD are well described. Omega-3 is known to be rich especially in marine fish. Recently, researchers reported the plant perilla mint (*Perilla frutescens*) as another potential source of this fatty acid. Particularly, the seeds of this plant have been reported with high omega-3 contents. When used as feed in swine and chicken, perilla seed and meal were found to increase omega-3 fatty acid contents in meat and other animal products such as eggs. An advantage of perilla mint is that production costs are much lower than for other crops such as soybean. Furthermore, perilla seeds have low levels of trypsin inhibitors, which make it suitable for use as animal feed.

**Keywords:** Perilla mint, carcass characteristic, meat quality

## Introduction

In 2013 and 2014 consumption rate of eggs in Thailand was 167.8 and 177.3 eggs per person per year, respectively. During 5 years (2010-2014) consumption rate of egg in Thailand increased by 4.77% annually (Office of Agricultural Economics, 2014), mainly due to the fact that eggs are available at lower costs than other protein sources. Hence, improving nutrition through the consumption of eggs is an important issue for good health. Meat and poultry meat are 5 to 15 folds lower in omega-3 contents than seafood. Pork consumption of Thai people is further and further increasing and in 2013 Thai people consumed 14.0 kg per person per year, which increased to 14.88 kg in 2014. Poultry meat consumption in Thailand 2013 was 15.5 kg per person per year, increased by 7.73% in 2014 to 16.7 kg (Office of Agricultural Economics, 2015). This emphasizes the importance of egg and meat consumption as important source of protein for humans. However, it can cause negative effects in humans because of several compounds such as saturated fatty acids, cholesterol, toxic contaminations, nitrate-nitrite preservatives and

antibiotic. For example, saturated fatty acids and cholesterol contained in eggs and meat are causative for several diseases in human health.

Nowadays, the quality of animal products, especially from a human health perspective, is of increasing importance. One reason is the steady increase of coronary artery disease (CAD), the most important cause of cardiovascular mortality worldwide, with over 4.5 million deaths occurring in the developing countries alone and 29% of Patients in Thailand were cardiovascular disease which is the highest than other disease (WHO, 2014). One of the most important risk factors for CAD is the nutrition. The fatty acid composition, especially those of the saturated fatty acids, represents a major risk factor. Consuming food rich in saturated fatty acids (i.e., meat, butter, and cheese) increases the amount of low density lipoprotein (LDL) in the blood but also increases high density lipoprotein (HDL the “good” cholesterol) and decreases triglycerides (Mozaffarian *et al.*, 2010). The main functional difference between HDL and LDL is where they transport cholesterol to. LDL takes cholesterol from liver to the cells while HDL carries cholesterol excess from the blood stream into the liver for removing. The main structural

difference is that LDL particle is 50% cholesterol and 25% protein while an HDL particle is 20% cholesterol and 50% protein (Harvey and Ferrier, 2011). This in turn carries cholesterol to tissues, including the heart arteries, and is thus causative for atherosclerosis (LDL forms plaque in vessels and blocks the blood system) causing CAD (Sudheendran *et al.*, 2010). The main sources of saturated fatty acids in the food are animal products, such as milk, meat, and eggs. Also, several plant products such as chocolate and cocoa butter, coconut, and palm kernel oil are rich in saturated fatty acids. Consuming too much saturated fatty acid lead to CAD, heart stroke, heart disease, blood pressure, type 2 diabetes and many other diseases (de Souza *et al.*, 2015). Numerous previous studies reported positive effects of unsaturated fatty acids, namely  $\alpha$ -linolenic acid (omega-3) and linoleic acid (omega-6) (Lecarf, 2009), on human health by decreasing the risk for CAD and other diseases. Therefore, consumers worldwide are increasingly demanding food products that are rich in omega-3 fatty acids. Increased levels of omega-3 in eggs were realised by supplementation or adding several sources of omega-3 such as flaxseed, fish oil and microalgae (Fraeye *et al.*, 2012) in laying-hen diets. Thus, enhancing omega-3 levels in eggs and meat though the use of agricultural by-products such as perilla meal, which is rich in omega-3, may be one of the possibilities to produce healthy food for the consumer. It is well known people in Thailand consume rice are main course, that is different from westerner, their main course is meat. Therefore, meat enhance with omega-3 is an alternative for meat lover in Thailand and other countries which are like to meat.

Perilla mint is known by many names, including Chinese basil, purple mint, rattlesnake weed and beefsteak plant (Glenn *et al.*, 2010). The

plant can be found on pastures in Southern India, South Korea, northern Thailand, China, Japan, Vietnam and Taiwan (Hyun *et al.*, 2014). Perilla mint was found in the northern part of Thailand such as Chiang Mai, Chiang Rai, Phayao and Mae Hong Son, cultivated area are 1,360 acre can produce perilla seed 272,000 kg (Chairaungyost, 2012). As a traditional oilseed, it is belonging to the Family lamiaceae. It is widely used in households (CSIR, 1966) for sprinkle on top of soups, salads, sushi and pickles or garnishes (Li *et al.*, 2008; Meng *et al.*, 2009; Ha *et al.*, 2012). The plant is an annual mint and blooms once a year in cool season, thus only one annual harvest is possible (Ampanchai *et al.*, 2008). The different parts of the plant are used for different purposes: leaves are known for their antioxidant and phenolic compounds, while seeds are rich in protein and fat. The tribal population in northeast India have been consuming perilla seed as well as oil without any negative effects being reported (Longvah *et al.*, 2000).

In the following, this article reviews the present knowledge on perilla mint when used as animal feed in order to improve the quality of meat and other animal products in terms of increasing the omega-3 fatty acid content.

#### Chemical composition of *Perilla frutescens*

As generally in plants, the different parts of perilla mint differ largely in their chemical composition. The protein and fat content of its seeds was 18.1 and 40.1%, respectively (Joshi *et al.*, 2015). Longvah and Deosthale (1991) reported similar values (protein 17%, fat 51%). Most vegetable oils are good sources of linoleic acid, but only few vegetable oils contain significant amounts of  $\alpha$ -linolenic acids. Among them, perilla oil which have the highest contents of  $\alpha$ -linolenic acid (omega-3),

amounting nearly 57% (Longvah and Deosthale, 1991), linoleic acid (omega-6) 14% and oleic acid (omega-9) ranging from 14-23% (Asif, 2011). Contents are even higher than in fish oil (Kim and Choi, 2005 ). In addition, the content of saturated fatty acids, namely palmitic acid (5-7%) and stearic acid (0-1%) is lower than in other sources of oil (Table 1). When taking the FAO/WHO/UNU (1985) pattern of essential amino acid requirement for infants as the reference (Table 2), the protein scores of egg and cow milk are 100, while perilla whole seed and perilla without hull are 63 and 66, respectively. The amino acid lysine was the limiting amino acid in both cases. The amino acid score of perilla whole seeds and kernel were comparable to sunflower (61) and peanut (69), both of which also have lysine as the limiting amino acid (Bodwell and Hopkins, 1985). Consequently, perilla seeds can be considered as a better source of essential amino

acids than soybean seed or sunflower meal (Table 2).

Perilla meal is a by-product of perilla seed after defatting. Protein contents are about 40.1% with 39% of the total content being essential amino acids. These levels are somewhat lower than in most animal products (beef 52, egg 55 and cow milk 54%) (Longvah and Deosthale, 1998). Perilla meal has compared to other plants such as soybean meal, sunflower meal, canola meal, cotton seed meal and sesame meal similar protein levels. It is therefore a good protein source for animal diets and good for eliminating agricultural by-products (Kusanteay and Uriyapongson, 2016).

#### Benefits of omega-3 fatty acids on human health

Mono- and polyunsaturated fatty acids are also known as omega fatty acids. Omega-3 fatty acids are polyunsaturated fatty acids where the first

**Table 1. Fatty acid composition from different sources of oil (% of fatty acid)**

Fatty acid	Beef tallow	Corn oil	Perilla seed	Perilla oil	Fish oil	Sunflower oil	Soybean oil	Palm oil
<b>Saturated fatty acid</b>								
Myristic (C14:0)	3.96	-	-	-	-	-	-	0.88
Palmitic (C16:0)	28.36	11.85	5-7	6.80	25.66	5.73	12.0	46.16
Stearic (18:0)	20.87	-	0-1	2.07	6.65	3.47	5.0	5.07
Arachidic (20:0)	-	0.54	-	-	-	0.54	1.0	0.41
<b>Unsaturated fatty acid</b>								
Palmitoleic (16:1)	2.79	-	-	-	5.50	0.02	-	0.13
Oleic (18:1)	44.02	28.94	NA	15.23	13.82	26.45	19-34	37.18
Linoleic (18:2)(n-6)	-	57.29	14	2.07	1.62	62.33	48-60	9.08
$\alpha$ -linolenic (18:3) (n-3)	-	0.93	57	61.30	1.08	-	2-10	0.36
Eicosapentaenoic acid (20:5) (n-3)	-	-	NA	-	32.72	-	-	-
Docosahexaenoic acid (22:6)(n-3)	-	-	NA	-	6.47	-	-	-
Source	Kim <i>et al.</i> (2005)	Kim and Choi (2005)	Peiretti (2011)	Kim <i>et al.</i> (2005)	Kim and Choi (2005)	Rosa <i>et al.</i> (2009)	Su <i>et al.</i> (2015)	Su <i>et al.</i> (2015)

Table 2. Essential amino acids from different sources (mg/g protein)

Essential amino acid	EAA recommendati on for human (FAO/WHO/UN U, 1985)	Perilla seed	Soybean seed	Sunflower meal	Peanut meal	Egg	Cow milk
Histidine	22	31	12	8.8	6	22	27
Isoleucine	18	41	21.6	15.2	8.3	54	47
Leucine	25	62	36.2	21.9	15.3	86	95
Lysine	22	37	29.6	14.6	8.5	70	78
Methionine+	29	26	13.2	15.9	5.9	57	33
Cysteine							
Phenylalanine+	34	55	40	25.2	21.9	93	102
Tyrosine							
Threonine	30	34	19.3	12	8.1	47	44
Tryptophan	7	11	6.5	-	2.3	17	14
Valine	31	35	22.2	17.5	9.9	66	64
Source	FAO/WHO/UNU (1985)	Longvah and Deosthale (1998)	ENV/JM/MONO (2001)	Rosa <i>et al.</i> (2009)	Settaluri <i>et al.</i> (2012)	Longvah and Deosthale (1998)	Longvah and Deosthale (1998)

double bond is located at the third carbon from the methyl end of the fatty acid chain. Marine animals are rich in the two long-chain omega-3 fatty acids docosahexaenoic acid (DHA; 22:6) and eicosapentaenoic acid (EPA; 20:5). Plants usually contain the omega-3 fatty acid,  $\alpha$ -linolenic acid (18:3) as omega-3. Several studies have reported that  $\alpha$ -linolenic acids and its derivatives are beneficial for the human health and fulfill essential physiological functions (Tinoco, 1982; Zollner, 1986; Crawford, 1987; Budowski, 1988; Neuringer *et al.*, 1988). The consumption is, for example, known to lower blood lipid concentration, reduce the risk for thrombosis, prevent Alzheimer's disease, decrease the risk for vascular disease and sudden death from ventricular fibrillation and tachycardia (Azin and

Behnood, 2014). Omega-3 fatty acids fulfill neuroprotective actions in Parkinson's disease and exhibit protective effects in Alzheimer's disease, too (Asif, 2011). Omega-3 fatty acids prevent the neurotoxin-induced decrease of dopamine that normally occurs. Therefore, the dopamine system that was disrupted will cause Parkinson's disease (Talbot and Hughes, 2006).  $\alpha$ -linolenic acid is the precursor of n-6 and n-3 polyunsaturated fatty acid (PUFA) families, linoleic acid (C18:2n-6) and  $\alpha$ -linolenic acid (C18:3n-3) are finally transformed to EPA and DHA in vivo (Weizhuo *et al.*, 2013). The ratio of linoleic acid and  $\alpha$ -linolenic acid in the diet is important for converting into long chain polyunsaturated fatty acids, including EPA and DHA (Diwakar *et al.*, 2008). As above-mentioned, these

affect the development of the human brain. Both EPA and DHA have several effects on inhibiting development of atherosclerosis, decrease triglyceride level and raise levels of HDL (high density lipoprotein “good cholesterol”) in blood plasma, but do not lower plasma cholesterol levels (Conner, 2001). Moreover, the impact of them may be primarily in preventing or relieving arrhythmia. Harris *et al.* (2009) reported cardiac mortality to be reduced by about 35% by modest EPA+DHA consumption. Although, omega-3 fatty acid, DHA and EPA have many advantages for the health but if consumed excessively (normally over 3 g per day), especially in patient that at the same time consume aspirin or warfarin, will increase bleeding or hemorrhagic stroke (only in cases of very large doses) and reduced glycemic control among diabetics (Lewis, 2008; Kromann and Green, 1980). According to the Food and Nutrition Board 2015 (Ornish, 2016) the acceptable intake for n-3 fatty acid is 1.6 g/day and 1.1 g/day for men and women, respectively (Asif, 2011).

#### Use of perilla mint as animal feed

Perilla mint was used as protein source but in their seed compound with a lot of oil. Fats and oils are a chemically diverse group of compounds. They have the highest average energy density among all macro nutrients. Therefore, fat is an important energy in diet and improve palatability and texture of diet in addition decrease dustiness and easy to make pellets. But they have limit level to use in animal diet such as not over 7% in swine diet because it causes flow ability problems preclude with swine (Shannon, 2016), in poultry not over 4% because it effect to decrease digestibility (Tancharoenrat, 2014). Perilla mint is not have only benefit but also have negative effect, some research reported about toxicity of

perilla mint especially, perilla's leaves is important problem in cattle poisoning that cause pulmonary edema (Brenner, 1993) because perilla ketone substance which found in leaves stem flower and root (Wilson *et al.*, 1977; Glenn *et al.*, 2010). Planta are most toxins when farmer cut and dried for hay late in summer, during flowering and seed production because flowering structure are the most dangerous but the ketone was not found in seed from perilla mint. Perilla ketone is not only harmful in ruminant but also toxic in small ruminant and mice (Glenn *et al.*, 2010).

Perilla mint was used as feed in animals for studying the mechanism of digestion or immunology. The common model species were rats or mice which are easy to manage and observe. Ihara *et al.* (1998) used the oil of safflower and perilla feed in male Wistar rats, 3-weeks old, and observed the lipid metabolism. Two diets either containing safflower oil (SO, high linoleic acid) or perilla oil (PO, high  $\alpha$ -linolenic acid) were compared. These diets were fed to rats for 3, 7, 20 and 50 days. The results showed that the level of cholesterol, triglycerides and total lipid in the PO-diet group were lower than in the SO-group feeding the diets for 7 days. This experiment demonstrated that  $\alpha$ -linolenic acid plays an important role in the regulation of serum cholesterol and the magnitude of regulation is more powerful when compared to linoleic acid.

Perilla mint has been used to feed animals because of its low costs and abundance throughout East Asia. Oita *et al.* (2008) studied the extraction and digestion of *Perilla frutescens*. In their experiment water-and NaCl-soluble protein fraction contained less trypsin inhibitors than that of soybeans.

As mentioned above, perilla seed and meal are suited as animal feed. Perilla seed was used in

polyunsaturated fatty acids (PUFA) enriched diets for growing rabbit aged of 73 days in order to increase the content of n-3 PUFA. Peiretti *et al.* (2010) fed perilla seed to rabbits at a proportion of 10% of the diet. Their results showed that neither digestibility of energy (DE) nor digestibility of protein (DP) and the ratio of DP/DE differed when compared with soybean meal. In during the growth stage of swine fattening, Thirty-one three-way crossbred Landrace×Large White×Duroc pigs, perilla meal was included into diets at 10, 15 and 20% (fresh matter). The 10%-group required the least amount of time to reach the target weight (110 kg) and showed the highest value for daily gain. In contrast, the control group without perilla showed the highest carcass percentages. The 15%-group showed medium back fat thickness. The 20%-group had a thinner back fat thickness of the shoulder than the other groups. Regarding the fatty acid composition of perineal fat, back fat, intermuscular fat and intramuscular fat positive correlations with increasing perilla meal proportions were found. The content of C18:3 increased, while the ratio of n-6/n-3 decreased. Furthermore, the ratio of C18:2/C18:0 and unsaturated fatty acid content also increased in a dose-dependent manner (Yamada *et al.*, 2007). Furthermore, Zang *et al.* (2003) added *Perilla frutescens* in laying hen diets at 8, 12, 16 and 20% (fresh matter), respectively. The results showed that the contents of PUFA increased to 20.42, 23.61, 24.07 and 24.62% in 8, 12, 16 and 20% supplemented groups, respectively, in contrast to 17.03% in the control group ( $P>0.05$ ). The content of omega-3 fatty acid in yolk increased to 6.88, 8.72, 9.86 and 9.95% in the correspondent groups, being significantly higher than in the control group (1.21%,  $P<0.01$ ). There was no significant difference in the total cholesterol (TCH). *Perilla frutescens* could be supplemented in the diet in order to increase the

content of PUFA and omega-3 fatty acid, improve the ratio of omega-6 to omega-3 and PUFA to SFA, and decrease TCH and the cost. Likewise, Saito *et al.* (2006) studied feeding perilla oil and seed meal in laying hens and observed  $\alpha$ -linolenic acid contents in yolk egg. The  $\alpha$ -linolenic acid contents of the egg yolks in perilla oil and perilla meal at the dietary levels of 0, 2.5, 5, 7.5 and 10% each were 0.20, 1.20, 1.64, 2.59 and 3.00 mg/100 mg egg yolk, and 0.17, 0.21, 0.28, 0.32 and 0.39 mg/100 mg egg yolk, respectively. It was suggested that perilla seeds may be the most suitable feed for the production of eggs with high  $\alpha$ -linolenic acid content from an economical point of view. Considering eggs are a rich source of dietary cholesterol and cardiovascular disease patient cannot eat egg over one egg per day (Zachary *et al.*, 2017). But if we improve fatty acid profile in egg by increase omega-3 it will be a good alternative for cardiovascular disease patient and general person because omega-3 can respect to primary and secondary prevention of coronary heart disease (Clayton *et al.*, 2015). So if they eat egg compound with omega-3 maybe reduce the risk cause cardiovascular disease in normal human and patient can eat egg by decrease concern.

Several researches indicated perilla seed and meal can improve fatty acid composition in meat and egg by increasing unsaturated fatty acid, especially omega-3 that has many benefits for human health. Using perilla mint in animal feed is still not pervasive to use in animal feed because the limitation of it such as fiber composition is quite high (problem with monogastric animal), cultivation in some area and harvest once time per year. It is interesting to use for increase value in meat and egg due to the property is high with protein and omega-3.

## Conclusion

Seeds of perilla mint, of which all parts of the plant can be used as animal feed, are rich in  $\alpha$ -linolenic acid. Through its use as feed, omega-3 fatty acid concentrations in meat and other animal products, i.e., eggs, can be increased, which in turn has positive effects on human health and is known to reduce the risk of different diseases such as coronary artery disease. Additionally, leaves are rich in various bioactive and phenolic compounds and seed meal can be used as protein source. Feeding perilla mint can minimize the contamination with methyl-mercury, which is caused by feeding fish meal. Overall, the use of perilla mint as animal feed has many advantages from a human health perspective and is economically efficient.

## References

- Ampanchai, S., S. Nuglor, P. Puddhanon, P. Supapornhem and V. Champa. 2008. Effect of storage conditions on seed quality and oil content of *Perilla frutescens* (L.) Britt. in the upper north of Thailand. *Agricultural Science Journal* 39(3): 421-424.
- Asif, M. 2011. Health effects of omega-3,6,9 fatty acids: *Perilla frutescens* is a good example of plant oils. *Oriental Pharmacy and Experimental Medicine Journal* 11(1): 51-59.
- Azin, M.N. and B. Behnood. 2014. Omega-3 Supplements and Cardiovascular Diseases. *National Research Institute of Tuberculosis and Lung Disease* 13(1): 6-14.
- Bodwell, C. E. and D. T. Hopkins. 1985. Nutritional characteristics of oilseed proteins. pp. 221-257. *In: A.M. Altschul and H.L. Wilcke* (eds.). *New Protein Foods*. Volume 5. Seed Storage Protein. Academic Press, New York.
- Brenner, D.M. 1993. Perilla: botany, uses, and genetic resources. pp. 322-328. *In: J. Janick and J.E. Simon* (eds.). *New Crops*. John Wiley and Sons, NY.
- Budowski, P. 1988.  $\omega$ -3 fatty acids in health and disease. *World Review of Nutrition and Dietetics* 57: 214-274.
- Clayton, Z.S., E. Fusco and M. Kern. 2017. Egg consumption and heart health: A review. *Nutrition* 37: 79-85.
- Connor, W. E. 2001. n-3 Fatty acids from fish and fish oil: panacea or nostrum?. *The American Journal of Clinical Nutrition* 74(4): 415-416.
- Council of Scientific Industrial Research (CSIR). 1966. *The Wealth of India: Raw Materials*. Publications and Information Directorate, Council of Scientific and Industrial Research, New Delhi.
- Crawford, M. A. 1987. The requirements of long chain n-6 and n-3 fatty acids for the brain. pp. 270-295. *In: W.E.M. Land* (ed.). *Proceedings of the AOCS short course on polyunsaturated fatty acids and eicosanoids*. American Oil Chemists Society, Champaign, Illinois.
- de Souza, R., A. Mente, A. Maroleanu, A. Cozma, V. Ha, T. Kishibe, E. Uleryk, P. Budylowski, H. Schünemann, J. Beyene and S.S. Anand. 2015. Intake of saturated and trans unsaturated fatty acids and risk of all cause mortality, cardiovascular disease, and type 2 diabetes: systematic review and meta-analysis of observational studies. *British Medical Journal* 351: h3978.



- Diwakar, B.T., P.K. Dutta, B.R. Lokesh and K.A. Naidu. 2008. Bio-availability and metabolism of n-3 fatty acid rich garden cress (*Lepidium sativum*) seed oil in albino rats. Prostaglandins Leukot Essent Fatty Acids 78(2): 123-30.
- ENV/JM/MONO. 2001. Consensus document on compositional considerations for new varieties of soybean: key food and feed nutrients and anti-nutrients. Series on the Safety of Novel Foods and Feeds 5(9): 1-60.
- FAO/WHO/UNU. 1985. Joint Expert Consultation. Energy and Protein Requirements. Technical Report Series 724. WHO, Geneva.
- Fraeye, I., C. Bruneel, C. Lemahieu, J. Buyse, K. Muylaert and I. Foubert. 2012. Dietary enrichment of eggs with omega-3 fatty acids: A review. Food Research International Journal 48: 961-969.
- Gester, H. 1998. Can adults adequately convert alpha-linolenic acid (18:3n-3) to eicosapentaenoic acid (20:5n-3) and docosahexaenoic acid (22:6n-3)?. International Journal of Vitamin and Nutrition Research 68: 159-173.
- Glenn, N., J. Bill and S. Brad. 2010. Beefsteake Plant (*Perilla* mint). Purdue Weed Science. Ws-043-w.
- Ha, T.H., J.H. Lee, M.H. Lee, B.W. Lee, H.S. Kwon, C.H. Park, K.B. Shim, H.T. Kim, I.Y. Baek and D.S.Jang. 2012. Isolation and identification of phenolic compounds from the seeds of *Perilla frutescens* (L.) and their inhibitory activities against  $\alpha$ -glucosidase and aldose reductase. Food Chemistry 135: 1317-1403.
- Harris, W.S., D. Mozaffarian, M. Lefevre, C.D. Toner, J. Colombo, S.C. Cunnane, J.M. Holden, D.M. Klurfeld, M.C. Morris and J. Whelan. 2009. Towards establishing dietary reference intakes for eicosapentaenoic and docosahexaenoic acids. Journal of Nutrition 139(4): 804S-819S (supplement).
- Harvey R.A. and D.R. Ferrier. 2011. Lippincott's Illustrated Reviews: Biochemistry. 5<sup>th</sup> ed. Lippincott Williams and Wilkins, Philadelphia. 531 p.
- Hyun, J., T.K. Beom, S.S. Geun and S.K. Young. 2014. Structural characterization of phenolic antioxidants from purple perilla (*Perilla frutescens* var. *acuta*) leaves. Food Chemistry. 148: 267-272.
- Ihara, M., H. Umekawa, T. Takahashi and Y. Furuichi. 1998. Comparative effects of short- and long-term feeding of safflower oil and perilla oil on lipid metabolism in rats. Comparative Biochemistry and Physiology Part B 121: 223-231.
- Joshi, A., A. Sharma, D. P. Pandey and R. K. Bachheti. 2015. Physico-chemical properties of *Perilla frutescens* seeds. Der Pharma Chemica 7(5): 35-41.
- Kromann, N. and A. Green. 1980. Epidemiological studies in the Upernavik district, Greenland. Incidence of some chronic diseases 1950-1974. Acta Medica Scandinavica 208(5): 401-406.
- Kusanteay, D. and S. Uriyapongson. 2016. Effect of dried tomato pomace in concentrate diets on nutrient digestibility and growth performance of Thai native cattle. Journal of Agriculture 32(2): 261-271.
- Lecerf, M.J. 2009. Fatty acids and cardiovascular disease. Nutrition Reviews 67(5): 273-83.

- Lewis, C.J. 2008. Letter regarding dietary supplement health claim for omega-3 fatty acids and coronary heart disease. (Online). Available: [http://deanornish.com/wp-content/uploads/the\\_Dark\\_Side\\_of\\_Good\\_Fats\\_Deans\\_Ornish.pdf?b70fc9](http://deanornish.com/wp-content/uploads/the_Dark_Side_of_Good_Fats_Deans_Ornish.pdf?b70fc9) (October 21, 2016).
- Li, Q., G. Yan and T. Ge. 2008. A fragmentation study of two compounds related to 4'-demethylepipodophyllotoxin in negative ion electrospray ionization by MSn ion-trap time-of-flight mass spectrometry. *Rapid Communication in Mass Spectrometry* 22(3): 373-378.
- Longvah, T. and Y.G. Deosthale.1991. Chemical and nutritional studies on Hanshi (*Perilla frutescens*) a traditional oilseed from northeast India. *Journal of the American Oil Chemists Society* 68: 781-784.
- Longvah, T. and Y.G. Deosthale.1998. Effect of dehulling, cooking and roasting on the protein quality of *Perilla frutescens* seed. *Food Chemistry* 63(4): 519-523.
- Longvah, T., Y.G. Deosthale and K. P. Uday. 2000. Nutritional and short term toxicological evaluation of perilla seed oil. *Food Chemistry* 70(1): 13-16.
- Meng, L., Y. Loazno, E. Gaydou and B. Li. 2009. Antioxidant activities of polyphenols extracted from *Perilla frutescens* varieties. *Molecules* 14(1): 133-140.
- Mozaffarian, D., R. Micha and S. Wallace. 2010. Effects on coronary heart disease of increasing polyunsaturated fat in place of saturated fat: A systematic review and meta-analysis of randomized controlled trials. *PLOS Medicine Journal* 7(3): e1000252.
- Nestel, P., P. Clifton, D. Colquhoun, T.A. Mori and B. Thomas. 2015. Indications for omega-3 long chain polyunsaturated fatty Acid in the prevention and treatment of cardiovascular disease. *Heart, Lung and Circulation* 24(8): 769-779.
- Neuringer, M., G.J. Anderson and W.E. Lonnor. 1988. The essentiality of n-3 fatty acids for the development and function of the retina and brain. *Annual Review of Nutrition* 8: 517.
- Office of Agricultural Economics. 2014. Important situation of agriculture product and tendency in 2015 (chicken). Office of Agricultural Economics, Bangkok.
- Office of Agricultural Economics. 2015. Important situation of agriculture product and tendency in 2015 (swine). Office of Agricultural Economics, Bangkok.
- Oita, S., T. Kimura, Y. Shibuya, N. Nihei and K. Tanahashi. 2008. Extraction and digestibility of *Perilla frutescens* seed protein. *Japan Agricultural Research Quarterly* 42(3): 211-214.
- Ornish, D. 2016. The dark side of good fats in newweek. (Online). Available: <:///C:/Users/Michael/Documents/Focus97/F9..otes/The%20Dark%20Side%20of%20Good%20Fats.htm> (December 19, 2016).
- Peiretti, P.G., F. Gai, G. Meineri, I. Zoccarato and L. Gasco. 2010. Apparent digestibility of compound diets with increasing levels of perilla (*Perilla frutescens* L.) seeds in rabbit. *Italian Journal of Animal Science* 9(81): 425-428.
- Peiretti, P. 2011. Fatty acid content and chemical composition of vegetative parts of perilla (*Perilla frutescens* L.) after different growth

- lengths. Research Journal of Medicinal Plants 5(1): 72-78.
- Rosa, P., R. Antoniassi, S. Freitas, H. Bizzo, D. Zanotto and M. Oliveira. 2009. Chemical composition of Brazilian sunflower varieties. Helia Journal 32(50): 145-155.
- Saito, K., M. Nomura and J. Kimura. 2006. Effect of feeding perilla oil, meal and leaves on fatty acid contents in egg yolk. (Online). Available: <http://agris.fao.org/agris-search/search.do?recordID=JP2002002526> (December 1, 2016).
- Settaluri, V.S.K., C.V.K. Kandala, N. Puppala and J. Sundaram. 2012. Peanuts and their nutritional aspects-a review. Food and Nutrition Sciences 3(12): 1644-1650.
- Shannon, M. 2016. Which fat source should I feed my pigs? (Online). Available: <http://swine.missouri.edu/nutrition/pigfatsource.htm> (December 16, 2016).
- Su, Y., Y. She, Q. Huang, C. Shi, Z. Li and C. Huang. 2015. The effect of inclusion level of soybean oil and palm oil on their digestible and metabolizable energy content determined with the difference and regression method when fed to growing pigs. Asian-Australasian Journal of Animal Science 28(12): 1751-1759.
- Sudheendran, S., C.C. Chang and R.J. Deckelbaum. 2010. N-3 vs. saturated fatty acids: Effects on the arterial wall. Prostaglandins Leukot Essent Fatty Acids 82(4-6): 205-209.
- Talbott, S.M. and K. Hughes. 2006. In The Health Professional's Guide to Dietary Supplements. Lippincott Williams and Wilkins, Baltimore, MD.
- Tancharoenrat, P. 2014. Factors considered in using fat in broilers feed formulation. Thai Journal of Animal Science 1(3): 7-16.
- Tinoco, J. 1982. Dietary requirements and functions of  $\alpha$ -linolenic acid in animals. Progress in Lipid Research 21(1): 1-45.
- Weizhuo, T., S. Yanling, L. Yanze, W. Yingfeng and Z. Yqing. 2013. Fast extraction of bioactive fatty acids from the perilla seeds by SMASH tissue extraction. Pakistan Journal of Botany 45(2): 513-517.
- WHO (World Health Organization). 2014. Noncommunicable Diseases Country Profiles 2014. WHO Document Production Services Publishing, Geneva. 201 p.
- Wilson, B.J., J.E. Garst, R.D. Linnabary and R.B. Channell. 1977. Perilla ketone: a potent lung toxin from the mint plant, *Perilla frutescens*. Britton Science 197(4303): 573-574.
- Yamada, M., A. Soeta, S. Sekiguti, J. Aminaka, K. Yamada and K. Mutou. 2007. Effects of *Perilla* meal feeding on the growth, carcass characteristics, and fatty acid composition of fat tissue and muscle in fattening pigs. Japanese Journal of Swine Science 44(2): 45-53.
- Zang, S., T. Li, Y. Song and X. Wang. 2003. Effects of *Perilla frutescens* in diet of laying hens on polyunsaturated fatty acid (PUFA) in egg yolk. Animal Husbandry & Veterinary Medicine 35(6): 13-16.
- Zollner, N. 1986. Dietary  $\alpha$ -linolenic acid in man-an overview. Progress in Lipid Research 25: 177-180.