Pollen Morphology of the Genus *Campylotropis* (Leguminosae) in Thailand and Its Systematic Implications

JIRATTHI SATTHAPHORN¹, WONGKOT PHUPHUMIRAT² AND CHARAN LEERATIWONG¹*

¹Department of Biology, Faculty of Science, Prince of Songkla University, Hatyai, Songkhla, 90110, THAILAND

²Department of Applied Science, Faculty of Science, Prince of Songkla University,

Hatyai, Songkhla, 90110, THAILAND

* Corresponding author. Charan Leeratiwong (charan.leeratiwong@gmail.com)
Received: 23 December 2019; Accepted: 19 May 2020

ABSTRACT.— Pollen grains of seven species of the genus *Campylotropis* in Thailand were described and illustrated based on investigations by light microscopy (LM) and scanning electron microscopy (SEM). The pollen grains were found to be generally monad, isopolar, radially symmetric and prolate. Apertures were tricolporate with lalongate endoapertures. The margo along colpi was psilate, perforate or absent. Exine ornamentation was microreticulate or microreticulate to reticulate, with lumina size diminishing toward the aperture. Types of lumina were a vertical or declined margin. The results of this study illustrated characteristics that can assist investigation of some *Campylotropis* pollen grains at interspecific level: presence and type of margo, type of ornamentation and type of lumina margin. Palynological studies of *Campylotropis bonii*, *C. harmsii* and *C. pinetorum* are firstly reported here. In addition, palynological description of each species, photographs and identification key to species based on palynological evidence are also presented.

KEY WORDS: Campylotropis, Leguminosae, palynology, Papilionoideae, Thailand

INTRODUCTION

The genus Campylotropis Bunge (Leguminosae, Papilionoideae) comprises about 37 species distributed in temperate and tropical Asia (Iokawa and Ohashi, 2002, 2008; Lewis et al., 2005; Puhua et al., 2010). The genus is morphologically distinguished by pinnately trifoliate leaves, light to dark violet, dark blue or pinkishwhite to creamy-white corolla and 1-jointed, 1-seeded pods. In Thailand, seven taxa are currently enumerated on the basis of comparative macro morphology viz. C. bonii Schindl., C. capillipes (Franch.) Schindl. subsp. prainii Iokawa and Ohashi, C. decora (Kurz) Schindl., C. harmsii Schindl., C. parviflora (Kurz) Schindl., C. pinetorum (Kurz) Schindl. and C. sulcata Schindl. (Satthaphorn et al., 2018).

The pollen morphology of the genus was first described by Ohashi (1971) followed by subsequent authors, Mitra and Mondal (1982), Perveen and Qaiser (1998) and Xu et al. (2011). The major palynological study of the genus was conducted by Iokawa and Ohashi (2002) based on 24 species. Pollen grains of Campylotropis were reported as having tricolporate aperture with psilate colpi. They were generally prolate, subprolate or prolate spheroidal in shape, isopolar and radially symmetric. Exine sculpturing was noted as fine to coarse reticulate (Mitra and Mondal, 1982; Iokawa and Ohashi, 2002), while Xu et al. (2011) also reported glabrate type in exine ornamentation. However, Campylotorpis species have not been delimited by palynological characteristics nor is there a key to species for identifying infrageneric taxa.

Although variations in pollen surface characteristics were recognized as valuable aids in delimiting taxonomic relationships (Van Den Berg, 1984; Rajbhandary et al., 2012), the knowledge of pollen morphology of *Campylotropis*, especially in Thailand, is inadequate due to insufficient sampling. This investigation of pollen flora of the Thai *Campylotropis* and its taxonomic significance aims to clarify details of variations and to construct the a key of pollen. A discussion on the delimitation of Thai *Campylotropis* proposes that some species have been palynologically unknown.

MATERIALS AND METHODS

The pollen grains of all Thai Campylotropis species were sampled from herbarium specimens cited in the description (herbarium acronym follows Thiers, 2020). Pollen samples were treated using the acetolysis method following Erdtman (1954). For examination with light microscopy (LM), pollen grains were mounted in glycerine jelly and observed with the Olympus BX43 (Zetter, 1989). The lengths of polar and equatorial axes were measured and the exine thickness including outline, shape and ornamentation were recorded. Measurements were taken from at least 20 grains for each sample and are given with the standard deviation (SD). For examination with scanning electron microscopy (SEM), pollen grains were directly taped onto the specimen stub. The grains were coated with gold (Au) on the stub using an SPI-MODULETM Sputter Coater, SPI 11425. SEM was conducted with the FEI Quanta 400 apparatus and the overview and ornamentation at the center of the equatorial point photographed. The palynological terminology is based on the criteria of Punt et al. (2007), Hesse et al. (2009) and Halbritter et al. (2018).

RESULTS

The results of palynological investigation of Thai Campylotropis are summarized in Table 1 and Figures 1-5. Pollen grains of Campylotropis were monad. isopolar, radially symmetric and prolate. The outline was convex triangular in polar view and elliptic in equatorial view. The length of the polar axis ranged from 19.8 µm (C. bonii) to 40.0 µm (C. harmsii) and the length of the equatorial axis from 15.6 µm (C. bonii) to 37.0 µm (C. harmsii). Apertures were tricolporate with lalongate endoapertures. The margo along colpi was psilate (C. bonii and C. sulcata) or perforate (C. capillipes subsp. prainii, C. decora, C. harmsii and C. parviflora), but absent in C. pinetorum. The length of colpi ranged from 15.4 µm (C. bonii) to 25.6 µm (C. decora). Protrusions at aperture were found in C. harmsii, C. parviflora and C. pinetorum, but were absent in the other species. Exine thickness ranged from 0.8 µm (C. pinetorum and C. sulcata) to 1.8 µm (C. bonii and C. decora). Exine ornamentation was microreticulate (C. bonii, C. decora, C. parviflora and C. pinetorum) and microreticulate to reticulate (C. capillipes subsp. prainii, C. harmsii and C. sulcata). Size of lumina was smaller towards the aperture. Margin of lumina was vertical in most species except declined in C. capillipes subsp. prainii and C. decora. Lumina diameter ranged from 0.2 µm (C. decora) to 2.0 µm (C. harmsii) with perforation at the base. The width of muri ranged from 0.2 µm (C. bonii, C. capillipes subsp. prainii and C. pinetorum) to 0.6 µm (C. decora, C. parviflora and C. sulcata). The identification key of Thai Campylotropis presented below is based on pollen morphology and a description of each species is appended.

TABLE 1. Investigation of pollen grains of *Campylotropis* in Thailand

Taxa	Polar axis (μm) min-max (average)	Equatorial axis (µm) min–max (average)	Colpi length (µm)	Margo	Orna- mentation type	Lumina margin	Lumina width (µm)	Muri width (µm)	Exine thickness (μm)	Voucher specimens
C. bonii	19.8–24.5 (22.4±1.2)	15.6–21.3 (19.3±1.2)	15.4–18.2	psilate	micro- reticulate	vertical	0.4-0.6	0.2-0.4	1.0–1.8	Satthaphorn and Leeratiwong 81 (PSU)
C. capillipes subsp. prainii	24.5–30.0 (27.5±1.7)	19.6–26.3 (23.9±1.5)	17.3–21.1	perforate	micro- reticulate to reticulate	declined	0.5–1.1	0.2-0.5	0.9–1.3	Satthaphorn and Leeratiwong 90 (PSU)
C. decora	22.5–32.5 (29.1±2.1)	19.6–27.5 (24.8±2.0)	20.6–25.6	perforate	micro- reticulate	declined	0.2-0.9	0.4-0.6	1.1–1.8	Satthaphorn and Leeratiwong 91 (PSU)
C. harmsii	25.4-40.0 (31.7±3.7)	20.7–37.0 (27.6±3.6)	22.8–24.5	perforate	micro- reticulate to reticulte	vertical	0.8–2.0	0.3-0.5	1.0–1.2	Satthaphorn and Leeratiwong 88 (PSU)
C. parviflora	25.0–30.0 (27.8±1.5)	18.4–23.8 (21.8±1.3)	20.9–22.9	perforate	micro- reticulate	vertical	0.3-0.9	0.3-0.6	0.9–1.3	Satthaphorn and Leeratiwong 92 (PSU)
C. pinetorum	23.8–29.5 (26.2±1.7)	19.5–25.0 (22.3±1.5)	19.5–21.6	absent	micro- reticulate	vertical	0.4-0.9	0.2-0.3	0.8–1.3	Satthaphorn and Leeratiwong 84 (PSU)
C. sulcata	22.4–28.5 (24.8±1.6)	17.9–24.5 (21.2±1.6)	18.9–22.0	psilate	micro- reticulate to reticulate	vertical	0.3–1.3	0.4-0.6	0.8–1.5	Satthaphorn 75 (PSU)

Key to the species based on pollen morphology

1. C. bonii
7. C. sulcata
3. C. decora
<i>lipes</i> subsp. <i>prainii</i>
5. C. parviflora
4. C. harmsii
6. <i>C. pinetorum</i>

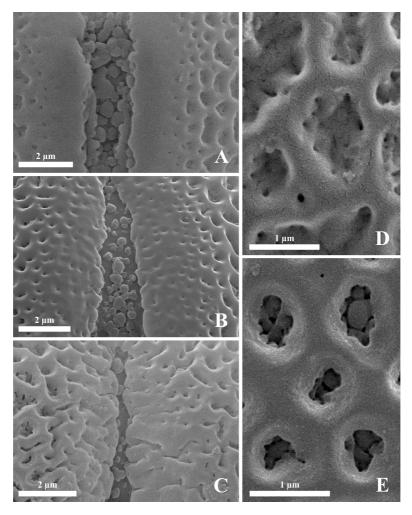


FIGURE 1. Significant characters that identify *Campylotropis* species in Thailand: A, psilate margo and absence of protrusion (*C. bonii*); B, perforate margo (*C. capillipes* subsp. *prainii*); C, absence of margo (*C. pinetorum*); D, vertical margin of lumina (*C. harmsii*); E, declined margin of lumina (*C. capillipes* subsp. *prainii*)

Pollen description of Thai Campylotropis

Pollen grains monad, isopolar, radially symmetric prolate; outline convex triangular in polar view, elliptic in equatorial view; tricolporate, margo psilate, perforate or absent; endoaperture lalongate; exine ornamentation microreticulate or microreticulate to reticulate; lumina margin vertical or declined, base perforate.

1. *Campylotropis bonii* (Figs. 2A–2D)

Description.— Length of polar diameter 19.8–24.5 μm; length of equatorial diameter 15.6–21.3 μm; margo psilate, colpi 15.4–18.2 μm long; endoaperture 4.5–5.6 μm long, 6.2–7.8 μm wide; exine 1.0–1.8 μm thick, microreticulate; lumina margin vertical, 0.4–0.6 μm wide with perforation, muri 0.2–0.4 μm wide.

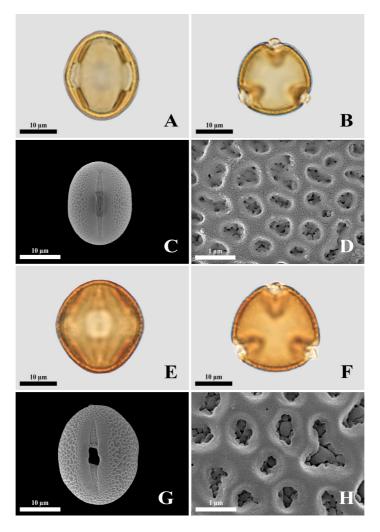


FIGURE 2. Pollen morphology of Thai *Campylotropis*: A-D, *C. bonii*; E-H, *C. capillipes* subsp. *prainii*; A & E, equatorial view; B & F, polar view; C & G, SEM overview of grains; D & H, SEM showing the ornamentation

Voucher specimen.— Satthaphorn and Leeratiwong 81 (PSU).

Note. — Pollen grains of *C. bonii* are resembled to pollen grains of those *C. sulcata* in having a psilate margo along colpi but exine sculpturing is different. Microreticulate ornamentation can be found in *C. bonii*, whereas *C. sulcata* exhibits microreticulate to reticulate ornamentation.

2. *Campylotropis capillipes* subsp. *prainii* (Figs. 2E–2H)

Description.— Length of polar diameter 24.5–30.0 μm; length of equatorial diameter 19.6–26.3 μm; margo perforate, colpi 17.3–21.1 μm long; endoaperture 6.6–7.4 μm long, 7.7–8.0 μm wide; exine 0.9–1.3 μm thick, microreticulate to reticulate; lumina

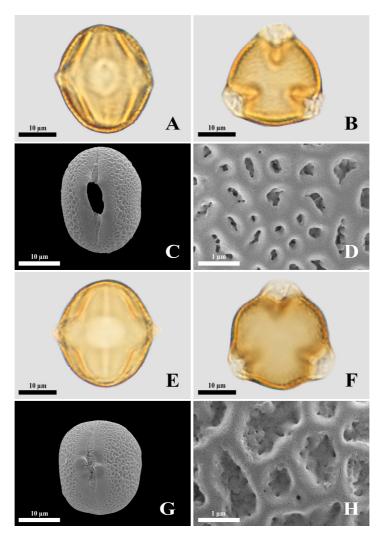


FIGURE 3. Pollen morphology of Thai *Campylotropis*: A-D, *C. decora*; E-H, *C. harmsii*; A & E, equatorial view; B & F, polar view; C & G, SEM overview of grains; D & H, SEM showing the ornamentation

margin declined, 0.5–1.1 μm wide, muri 0.2–0.5 μm wide.

Voucher specimen.— Satthaphorn and Leeratiwong 90 (PSU).

Note.— Pollen grains of the species exhibit the declined margin of lumina which is similar to pollen grains of *C. decora*. However, this character is different from the vertical margin of lumina in the other species.

3. *Campylotropis decora* (Figs. 3A–3D)

Description.— Length of polar diameter 22.5–32.5 μm; length of equatorial diameter 19.6–27.5 μm; margo perforate, colpi 20.6–25.6 μm long; endoaperture 8.0–10.4 μm long, 9.7–10.6 μm wide; exine 1.1–1.8 μm thick, microreticulate; lumina margin declined, 0.2–0.9 μm wide, muri 0.4–0.6 μm wide.

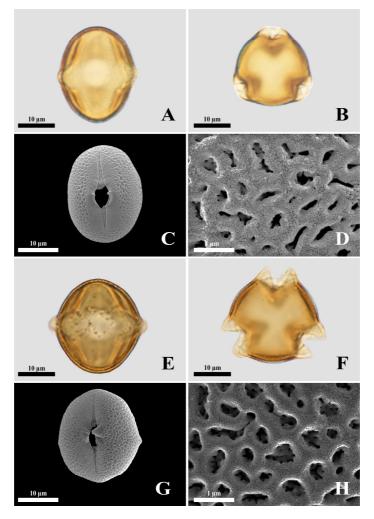


FIGURE 4. Pollen morphology of Thai *Campylotropis*: A-D, *C. parviflora*; E-H, *C. pinetorum*; A & E, equatorial view; B & F, polar view; C & G, SEM overview of grains; D & H, SEM showing the ornamentation

Voucher specimen.— Satthaphorn and Leeratiwong 91 (PSU).

4. *Campylotropis harmsii* (Figs. 3E–3H) **Description.**— Length of polar diameter 25.4–40.0 μm; length of equatorial diameter 20.7–37.0 μm; margo perforate, colpi 22.8–24.5 μm long; endoaperture 5.4–9.0 μm long, 7.6–9.0 μm wide; exine 1.0–1.2 μm thick, microreticulate to reticulate; lumina

margin vertical, $0.8-2.0 \mu m$ wide, muri $0.3-0.5 \mu m$ wide.

Voucher specimen.— *Satthaphorn and Leeratiwong* 88 (**PSU**).

5. Campylotropis parviflora (Figs. 4A–4D) **Description.**— Length of polar diameter 25.0–30.0 μm; length of equatorial diameter 18.4–23.8 μm; margo perforate, colpi 20.9–22.9 μm long; endoaperture 6.6–7.9 μm

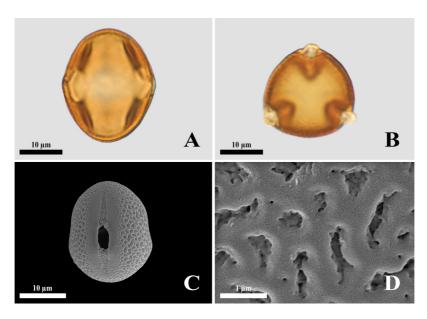


FIGURE 5. Pollen morphology of Thai *Campylotropis*: A-D, *C. sulcata*; A, equatorial view; B, polar view; C, SEM overview of grains; D, SEM showing the ornamentation

long, 7.4–8.9 μm wide; exine 0.9–1.3 μm thick, microreticulate; lumina margin vertical, 0.3–0.9 μm wide, muri 0.3–0.6 μm wide.

Voucher specimen.— *Satthaphorn and Leeratiwong 92* (**PSU**).

6. Campylotropis pinetorum (Figs. 4E–4H) **Description.**— Length of polar diameter 23.8–29.5 μm; length of equatorial diameter 19.5–25.0 μm; margo absent, colpi 19.5–21.6 μm long; endoaperture 8.0–8.6 μm long, 7.5–8.9 μm wide; exine 0.8–1.3 μm thick, microreticulate; lumina margin vertical, 0.4–0.9 μm wide, muri 0.2–0.3 μm wide.

Voucher specimen.— Satthaphorn and Leeratiwong 84 (PSU).

Note.— Pollen grains of *C. pinetorum* were apparently distinguished from other species by the absence of margo along the colpi.

7. Campylotropis sulcata (Figs. 5A–5D)

Description.— Length of polar diameter 22.4–28.5 μm; length of equatorial diameter 17.9–24.5 μm; margo psilate, colpi 18.9–22.0 μm long; endoaperture 6.7–7.9 μm long, 6.7–8.0 μm wide; exine 0.8–1.5 μm thick, microreticulate to reticulate; lumina margin vertical, 0.3–1.3 μm wide, muri 0.4–0.6 μm wide.

Voucher specimen.— Satthaphorn 75 (PSU).

DISCUSSION AND CONCLUSIONS

Pollen grains of the genus *Campylotropis* in this study are monad, prolate, isopolar, radially symmetric and tricolporate with lalongate endoaperture. This pollen morphology is congruent with palynological studies of Guinet (1981) and Ferguson and Skvarla (1981) that the basic type of pollen grains in the family Leguminosae is tectatereticulate single grains with tricolporate

aperture. This pollen type is recognized as an advance character in Angiosperms. Interestingly, flowers in the subfamily Papilionoideae have highly evolved with an explosive pollination mechanism which required pollinators such as insects, birds and bats, to stimulate pollen grains releasing (Aluri and Reddi, 1995; Miguel-Peñaloza et al., 2019). Ferguson and Skvarla (1982) revealed that entomophilous pollination species in the subfamily Papilionoideae contain simple reticulate or perforate surface sculpturing pollen grains, while birdpollinated and bats-pollinated species illustrate coarsely rugulate or verrucate pollen grains and stickier than those entomophilous pollen grains. Because the Campylotropis flowers are not showy and easily breaking, the pollinators of the genus are predominant by insect groups especially bees rather than those huge and solid papilionaceous flowers, for Erythina or Mucuna species which require birds or bats (Graham and Tomb, 1977; Arrayo, 1981).

The findings of this study uncovered characteristics that can assist investigation of some Campylotropis pollen grains at interspecific level. These characteristics included presence and type of margo (psilate, perforate and absent, Figs. 1A-1C), type of exine ornamentation (microreticulate vs microreticulate to reticulate) and type of lumina margin (vertical vs declined, Figs. 1D-1E). Even though the presence of protrusion seemed to be observed as a distinct feature in C. harmsii, C. parviflora and C. pinetorum (Figs. 3G, 4C, 4G), the study of Kuang et al. (2012) pointed out that the presence or absence of this character depends on anther stage and acetolysis technique. This character should be excluded from taxonomic structures. Although pollen morphology of C. capillipes subsp. prainii.

C. decora, C. parviflora and C. sulcata were described in previous studies (Ohashi, 1971; Mitra and Mondal, 1982; Perveen and Qaiser, 1998; Iokawa and Ohashi, 2002), this study presented the first diagnoses of the pollen morphology of C. bonii, C. harmsii and C. pinetorum.

Our results indicated that pollen grains of each species of Thai Campylotropis showed non-significant variations of P-axis and E-axis values which might be correlated to stability at species level. Mitra and Mondal (1982) reported average values of P-axis and E-axis lengths of C. capillipes subsp. prainii and C. parviflora were slightly longer than those in the present study. Likewise, Iokawa and Ohashi (2002) illustrated longer values of measurements in C. sulcata except those of C. parviflora and decora were shorter than measurements. These differences might be connected to diverse meiotic abnormalities. ploidy or geographical variation (Aguilar-García et al., 2012; Storme et al., 2013). According to Satthaphorn et al. (2018), this palynological information can be also used taxonomic implication in some morphological-related species in Thailand. For instance, C. parviflora and C. pinetorum exhibit pinkish-white to creamy-white corolla color with green blotches on the standard while other species display dark blue or light to dark violet color. However, their pollen morphology is different by having perforate margo in C. parviflora but absence in C. pinetorum. Another example is that C. capillipes subsp. prainii and C. bonii similarly pose appressed hairs on their peduncles while their pollen grains have distinct characters to discriminate: psilate margo and vertical margin of lumina in C. bonii, whereas perforate margo and declined margin of lumina in C. capillipes subsp. prainii.

The pollen grains of Campylotropis were also morphologically similar to those of most taxa in the tribe Desmodieae, such pollen from genera Alvsicarpus, Dendrolobium, Desmodium, Lespedeza, Kummerowia, Ohwia and Phyllodium (Ferguson and Skvarla, 1981; Chen and Huang, 1993; Perveen and Qaiser, 1998; Iokawa and Ohashi, 2002; Xu et al., 2011; Ohashi al.. 2013: Saisorn et Chantaranothai, 2015). Chen and Huang (1993) indicated that pollen grains of the genus *Dendrolobium* of the subtribe Desmodiinae were closely related to pollen of the subtribe Lespedezinae (Campylotropis, Lespedeza and Kummerowia) due to having coarse reticulate tectum. Moreover, this type of tectum was later observed in the genera Phyllodium and Ohwia (Ye et al., 2011; Ohashi et al., 2013). The close relation deduced from the similarity in pollen sculpture was recently supported molecular evidence which suggested that the subtribe Lespedezinae is phylogenetically sister to subtribe the Desmodiinae (Jabbour et al., 2018). However, brief details of exine sculpturing may not be taken into account when examining pollen morphology at infratribal level because of the occurrence of convergent and divergent characters among angiosperm pollen grains (Chen and Huang, 1993; Halbritter and Hesse, 1995).

In addition to palynological characteristics shared within the tribe Desmodieae, the coarsely reticulate tectum and lalongate endoaperture of *Campylotropis* were also found in pollen grains of *Lespedeza* and *Kummerowia* in previous investigations (Ferguson and Skvarla, 1981; Ohashi et al., 1981; Chen and Huang, 1993). These findings might indicate a close relationship within the subtribe Lespedezinae. The similarities of pollen grains were also consistent with phylogenetic molecular

studies that suggested Campylotropis is sister to Lespedeza with nested Kummerowia (Han et al., 2010; Nemoto et al., 2010; Jabbour et al., 2018). Because of the close relationship between the three genera, the delimitation of Campylotropis from the other two by pollen morphology is obscure, particularly from the genus Lespedeza. Notwithstanding. Kummerowia can be distinguished by its additional funiform structure on the exine sculpturing concealing lumina, while absent in Campylotropis and Lespedeza pollen grains (Xu et al., 2011). Generic recognition might also contradict intergeneric classification based on studies of macromorphology. It should be noted that the macromorphological studies compared flowers (two flowers per subtended bract in Lespedeza and Kummerowia with one flower per subtended bract in Campylotropis) and habits (shrubby in Campylotropis and Lespedeza vs herbaceous in Kummerowia) clearly distinguished between these three genera (Iokawa and Ohashi, 2002; Lewis et al., 2005; Puhua et al., 2010).

In conclusion, this palynological investigation provided information of pollen morphology, description and identification key to species based on palynological evidence for Thai Campylotropis. Information of pollen morphology of C. bonii, C. harmsii and C. pinetorum are firstly reported in the present study. implications of pollen morphology can be used to solve taxonomic aspects for distinguishing some morphological-related species in the genus. Clarification of the relationship within and between the genus Campylotropis and its related genera requires more samples and more evidence, such as anatomical and molecular studies from wider floristic regions or monographic scale.

ACKNOWLEDGEMENTS

The authors would like to express an appreciation to Prof. Pranom Chantaranothai for the valuable guidance. We would also thank to Mr. Thomas Duncan Coyne for proving grammatical English. This research was financially supported by Science Achievement Scholarship of Thailand (SAST), Graduate School, Dissertation Funding for Thesis, Prince of Songkla University and Center of Excellence on Biodiversity (BDC-PG3-160013).

LITERATURE CITED

- Aguilar-García, S.A., Figueroa-Castro, D.M. and Castañeda-Posadas, C. 2012. Pollen morphology of *Pachycereus weberi* (Cactaceae). Plant Systematics and Evolution, 98: 1845–1850.
- Aluri, R. and Reddi, C. 1995. Explosive pollen release and pollination in flowering plants. Proceedings of the Indian National Science Academy, B61(4): 323–332.
- Arroyo, M.T.K. 1981. Breeding systems and pollination biology in Leguminosae. In: Polhill, R.M. and Raven, P.H. (Eds). Advances in Legume Systematics, Royal Botanic Gardens, Kew, pp. 723–769.
- Chen, S.-J. and Huang, T.-C. 1993. Pollen morphology of the tribe Desmodieae (Leguminosae) in Taiwan. Taiwania, 38: 67–89.
- Erdtman, G. 1954. An introduction to pollen analysis. Almquist & Wiksell, Stockholm. 239 pp.
- Ferguson, I.K. and Skvarla, J.J. 1981. The pollen morphology of the subfamily Papilionoideae (Leguminosae). In: Polhill, R.M. and Raven, P.H. (Eds). Advances in Legumes Systematics 1, Royal Botanic Gardens, Kew, p. 859–894.
- Ferguson, I.K. and Skvarla, J.J. 1982. Pollen morphology in relation to pollinators in Papilionoideae (Leguminosae). Botanical Journal of the Linnean Society, 84: 183–193.
- Graham, A. and Tomb, A.S. 1977. Palynology of *Eythrina* (Leguminosae: Papilionoideae): the subgenera sections, and generic relationships. Lloydia, 40: 413–435.
- Guinet, P. 1981. Comparative account of pollen characters in the Leguminosae. In: Polhill, R.M. and Raven, P.H. (Eds). Advances in Legume

- Systematics, Royal Botanic Gardens, Kew, pp. 789–799.
- Iokawa, Y. and Ohashi, H. 2002. A taxonomic study of *Campylotropis* (Leguminosae) I. Journal of Japanese Botany, 77(4): 179–222.
- Iokawa, Y. and Ohashi, H. 2008 Campylotropis (Leguminosae) of China, an Enumeration and Distribution. Journal of Japanese Botany, 83: 36– 59.
- Jabbour, F., Gaudeul, M., Lambourdiere, J., Ramstein, G., Hassanin, A., Labat, J.-N. and Sarthou, C. 2018. Phylogeny, biogeography and character evolution in the tribe Desmodieae (Fabaceae: Papilionoideae), with special emphasis on the new Caledonian endemic genera. Molecular Phylogenetics and Evolution, 118: 108–123.
- Han, J.E., Chung, K.-H., Nemoto, T. and Choi, B.-H. 2010. Phylogenetic analysis of eastern Asian and eastern North American disjunct *Lespedeza* (Fabaceae) inferred from nuclear ribosomal ITS and plastid region sequences. Botanical Journal of the Linnean Society, 164: 221–235.
- Halbritter, H. and Hesse, M. 1995. The convergent evolution of exine shields in angiosperm pollen. Grana, 34: 108–119.
- Halbritter, H., Ulrich, S., Grímsson, F., Weber, M., Zetter, R., Hesse, M., Buchner, R., Svojtka, M. and Frosch-Radivo, A. 2018. Illustrated Pollen Terminology. Springer International Publishing. 483 pp.
- Hesse, M., Halbritter, H., Zetter, R., Weber, M., Buchner, R., Frosch-Radivo, A. and Ulrich, S. 2009. Pollen terminology, an illustrated handbook. Springer Wien New York, Austria. 266 pp.
- Kuang, Y.-F., Kirchoff, B.K. and Liao, J.-P. 2012. Presence of the protruding oncus is affected by anther dehiscence and acetolysis technique. Grana, 51: 253–262.
- Lewis, G.P., Schrire, B., Mackinder, B. and Lock, M. 2005. Legumes of the World. The Bath Press, UK. 592 pp.
- Ohashi, H. 1971. A taxonomic study of the tribe Coronilleae (Leguminosae), with a special reference to pollen morphology. Journal of the Faculty of Science, University of Tokyo, sect. III, 11: 25–92.
- Ohashi, H., Polhill, R.H. and Schubert, B.G. 1981. Desmodieae. In: Polhill, R.M. and Raven, P.H. (Eds). Advances in Legume Systematics 1, Royal Botanic Gardens, Kew, p. 292–300.
- Ohashi, K., Nata, K. and Ohashi, H. 2013. Pollen Morphology of the Genus *Ohwia* (Leguminosae:

- Tribe Desmodieae). Journal of Japanese Botany, 88: 291–296.
- Miguel-Peñaloza, A., Delgado-Salinas, A. and Jiménez-Durán, K. 2019. Pollination biology and breeding system of *Desmodium grahamii* (Fabaceae, Papilionoideae): functional aspects of flowers and bees. Plant Systematics and Evolution, 305: 743–754.
- Mitra, K. and Mondal, M. 1982. Pollen morphology of exstipellate and stipellate Hedysareae (Leguminosae). Proceedings of the Indian National Science Academy, B48: 755–769.
- Nemoto, T., Yokoyama, J., Fukuda, T., Iokawa, Y. and Ohashi, H. 2010. Phylogeny of *Lespedeza* (Leguminosae) based on chloroplast *trnL-trnF* sequences. Journal of Japanese Botany, 85(4): 213–229.
- Perveen, A. and Quiser, M. 1998. Pollen flora of Pakistan – VIII Leguminosae (subfamily: Papilionoideae). Turkish Journal of Botany, 22: 73–91.
- Puhua, H., Ohashi, H. and Iowaka, Y. 2010. Camyplotropis. In: Wu, Z.Y, Raven, P.H. and Hong, D.Y. (Eds). Flora of China 17, Science Press, Missouri, Botanical Garden, USA, p. 292–302.
- Punt, W., Hoen, P.P., Blackmore, S., Nilsson, S. and Le Thomas, A. 2007. Glossary of pollen and spore terminology. Review of Palaeobotany and Palynology, 143: 1–81.
- Rajbhandary, S., Hughes, M. and Shrestha, K. 2012. Pollen morphology of *Begonia* L. (Begoniaceae) in Nepal. Bangladesh Journal of Plant Taxonomy, 19(2): 191–200.
- Saisorn, W. and Chantaranothai, P. 2015. Taxonomic Studies on the Genus *Phyllodium* Desv.

- (Leguminosae) in Thailand. Tropical Natural History, 15(1): 23–40.
- Satthaphorn, J., Roongsattham, P., Chantaranothai, P. and Leeratiwong, C. 2018. The genus *Campylotropis* Bunge (Luguminosae) in Thailand. Thai Forest Bulletin (Botany), 46(2): 38–50.
- Storme, N.D., Zamariola, L., Mau, M., Sharbel, T.F. and Geelen, D. 2013. Volume-based pollen size analysis: an advanced method to assess somatic and gametophytic ploidy in flowering plants. Plant Reproduction, 26(2): 65–81.
- Thiers, B. 2020, continuously updated. Index Herbariorum: A global directory of public Herbaria and associated staff. New York Botanic Garden's Virtual Herbarium. http://sweetgum.nybg.org./ih/. January 25, 2020.
- Van den Berg, R.G. 1984. Pollen characteristics of the genera of the Begoniaceae. Agricultural University Wageningen Papers, 83(9): 55–66.
- Xu, B., Gao, X.-F., Wu, N. and Zhang, L.-B. 2011. Pollen diversity and its systematics implications in *Lespedeza* (Fabaceae). Systematics Botany, 36(2): 352–361.
- Ye, B., Ohashi, H. and Ohashi, K. 2011. Pollen Morphology of the Genera *Dendrolobium* and *Phyllodium* (Leguminosae: Papilionoideae Tribe Desmodieae). Journal of Japanese Botany, 86: 333–349.
- Zetter, R. 1989. Methodik und Bedeutung einer routinemäßig kombinierten lichtmikroskopischen und rasterelektronenmikroskopischen Untersuchung fossiler Mikrofloren. Cour Forsch-Inst Senckenberg, 109: 41–50.